Mesenchymal stem cell migration is regulated by fibronectin through \( \alpha 5\beta 1 \)-integrin-mediated activation of PDGFR-\( \beta \) and potentiation of growth factor signals

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Summary

Cell migration during vascular remodelling is regulated by crosstalk between growth factor receptors and integrin receptors, which together coordinate cytoskeletal and motogenic changes. Here, we report extracellular matrix (ECM)-directed crosstalk between platelet-derived growth factor receptor (PDGFR)-\( \beta \) and \( \alpha 5\beta 1 \)-integrin, which controls the migration of mesenchymal stem (stromal) cells (MSCs). Cell adhesion to fibronectin induced \( \alpha 5\beta 1 \)-integrin-dependent phosphorylation of PDGFR-\( \beta \) in the absence of growth factor stimulation. Phosphorylated PDGFR-\( \beta \) co-immunoprecipitated with \( \alpha 5 \)-integrin and colocalised with \( \alpha 5\beta 1 \)-integrin in the transient tidemarks of focal adhesions. Adhesion to fibronectin also strongly potentiated PDGF-BB-induced PDGFR-\( \beta \) phosphorylation and focal adhesion kinase (FAK) activity, in an \( \alpha 5\beta 1 \)-integrin-dependent manner. PDGFR-\( \beta \)-induced phosphoinositide 3-kinase (PI3K) and Akt activity, actin reorganisation and cell migration were all regulated by fibronectin and \( \alpha 5\beta 1 \)-integrin. This synergistic relationship between \( \alpha 5\beta 1 \)-integrin and PDGFR-\( \beta \) is a fundamental determinant of cell migration. Thus, fibronectin-rich matrices can prime PDGFR-\( \beta \) to recruit mesenchymal cells at sites of vascular remodelling.

Key words: Platelet-derived growth factor receptor-\( \beta \), \( \alpha 5\beta 1 \)-Integrin, Fibronectin, Mesenchymal stem cell, Signalling, Migration

Introduction

Signalling through the receptor tyrosine kinase (RTK) platelet-derived growth factor receptor (PDGFR)-\( \beta \) is essential for the migration and differentiation of cells during vascular development (Yancopoulos et al., 2000; Betsholtz et al., 2001; Kinner et al., 2002; Lindblom et al., 2003; Ball et al., 2007; Andrae et al., 2008). Knockout of the genes encoding PDGFR-\( \beta \) or PDGFR-B in mice causes death during late embryonic stages from widespread microvascular bleedings caused by deficient mural cell recruitment (Lindahl et al., 1997; Hellström et al., 1999; French et al., 2008). PDGF-BB, the main growth factor ligand of PDGFR-\( \beta \), is a potent stimulant of smooth muscle cell (SMC) recruitment during neointimal hyperplasia following vascular injury (Andrae et al., 2008). PDGFR signalling has also emerged as a predominant pathway in recruitment of adult perivascular mesenchymal stem cells (MSCs), which play crucial roles in angiogenesis, wound repair and tissue regeneration (Ferrari et al., 1998; Abedin et al., 2004; Fiedler et al., 2004; Tepper et al., 2005; Ball et al., 2007).

Dimerisation of PDGFR-\( \beta \), induced by ligating growth factor dimers, stimulates autophosphorylation of specific tyrosine residues within its cytoplasmic domain (Kelly et al., 1991). PDGFR-\( \beta \) is mainly activated by PDGF-BB, but also by PDGF-DD and vascular endothelial growth factor (VEGF)-A (Fredriksson et al., 2004; Ball et al., 2007). Autophosphorylation provides docking sites for downstream signal transduction molecules (Kazlauskas and Cooper, 1989), especially phosphoinositide 3-kinase (PI3K), which mediates actin reorganisation and migration, phospholipase C\( \gamma \) (PLC\( \gamma \)), which stimulates cell growth and motility, and Src family tyrosine kinases, which influence cell proliferation (Heldin et al., 1998; Jiménez et al., 2000; Tallquist and Kazlauskas, 2004; Andrae et al., 2008). Signalling by RTKs such as PDGFR-\( \beta \) is not only regulated by growth factors but also by functional collaboration with integrins (Eliceiri, 2001; Yamada and Even-Ram, 2002; Giancotti and Tarone, 2003; Streuli and Akhtar, 2009). Integrins are \( \alpha \beta \) heterodimeric cell-surface receptors that act as a transmembrane link between extracellular matrix (ECM) ligands and the actin cytoskeleton. They direct inside-out and outside-in signalling that regulates numerous cellular responses, including survival, growth, migration and differentiation (Hynes, 1992; Danen and Sonnenberg, 2003; Askari et al., 2009). \( \beta 1 \)- and \( \alpha \beta 3 \)-integrins can influence PDGFR-\( \beta \) activity (Sundberg and Rubin, 1996; Schneller et al., 1997; Woodard et al., 1998; Borges et al., 2000; Minami et al., 2007; Amano et al., 2008; Zemskov et al., 2009), and integrin-linked kinase (ILK) can control SMC migration in response to PDGF (Esfandiarei et al., 2010). However, the molecular mechanisms underlying crosstalk between PDGFR-\( \beta \) and integrins, and how they coordinate cell recruitment, remain obscure.

We have discovered a fundamental ECM-specific receptor crosstalk mechanism that controls MSC migration. Adhesion to fibronectin, through \( \alpha 5\beta 1 \)-integrin, specifically induced MSC migration by activating PDGFR-\( \beta \) signalling in the absence of growth factor stimulation. Fibronectin also strongly potentiated growth-factor-mediated receptor activation in an \( \alpha 5\beta 1 \)-integrin-dependent manner. Phosphorylated PDGFR-\( \beta \) appeared in ruffles at the leading edge of migratory cells and transiently colocalised with \( \alpha 5\beta 1 \)-integrin in the tidemarks of focal adhesions. Corresponding focal adhesion kinase (FAK)-dependent PI3K activity, actin reorganisation, membrane ruffling and MSC migration all exhibited fibronectin- and \( \alpha 5\beta 1 \)-integrin-dependence. This synergistic relationship between \( \alpha 5\beta 1 \)-integrin and PDGFR-
β is thus a fundamental determinant of mesenchymal cell migration. Fibronectin-rich matrices might therefore act to prime PDGFR-β to recruit mesenchymal cells to sites of vascular remodelling.

Results

Adhesion to ECM induces PDGFR tyrosine phosphorylation

To evaluate how adhesion to ECM regulates PDGFR activation in MSCs, tyrosine phosphorylation of PDGFR-α and PDGFR-β was examined in serum-free conditions, after plating onto fibronectin, laminin, fibrillin-1 PF8 [an Arg-Gly-Asp (RGD)-containing fragment that engages the α5β1-integrin] (Bax et al., 2007; Cain et al., 2005), collagen type I or collagen type IV (all at 10 μg/ml) for 90 minutes. A human array for phosphorylated RTKs (Fig. 1A), containing 42 different immobilised anti-RTK antibodies, was utilised; this allowed the simultaneous relative quantification of tyrosine phosphorylation levels for both PDGFR-α (array coordinates C7 and C8) and PDGFR-β (array coordinates C9 and C10) in the same cell lysate. MSCs plated onto a BSA control substrate (basal) for 90 minutes produced no detectable immunoreactivity for any RTKs, but control reactivity against phosphorylated tyrosine was positive. When MSCs were plated onto fibronectin for 90 minutes, PDGFR-α and PDGFR-β tyrosine phosphorylation was detected. Fibronectin stimulation also induced tyrosine phosphorylation of other RTKs, notably epidermal growth factor receptor (EGFR), EphA7, Axl and c-Ret. When MSCs were plated onto fibrillin-1 PF8, collagen type I or collagen type IV for 90 minutes, PDGFR-β phosphorylation was induced at lower levels compared with that on fibronectin, but no PDGFR-α phosphorylation was detected. Fibrillin-1 PF8, and collagen types I and IV, also induced tyrosine phosphorylation of EGFR (array coordinates B1 and B2). By contrast, MSCs plated onto laminin for 90 minutes showed no detectable RTK phosphorylation, similar to that with MSCs plated onto BSA. The percentage of MSCs adhering to the different ECM substrates after 90 minutes was similar (supplementary material Fig. S1A), indicating that the

Fig. 1. ECM-induced tyrosine phosphorylation of PDGFR-β. (A) Human phosphorylated RTK arrays were used to examine ECM-induced RTK phosphorylation levels in MSC lysates. PDGFR-α (R-α; coordinates C7 and C8) and PDGFR-β (R-β; coordinates C9 and C10), respectively, were examined in MSC lysates, taken at 90 minutes, from BSA-coated wells or wells coated with 10 μg/ml fibronectin, laminin, fibrillin-1 PF8, collagen type I or collagen type IV. Each array was identically exposed to detection reagents and film. Quantitative analysis was evaluated by densitometry with phosphorylated PDGFR duplicate RTK spots, normalised against phosphotyrosine-positive control spots [coordinates (A1, A2), (A23, A24), (F1, F2), (F23, F24)], and is represented as the fold increase above that with BSA control substrate (±s.d.) for duplicate spots. A representative example of two independent experiments is shown for each array analysis. (B) Immunoprecipitation (IP) analysis of the levels of PDGFR-β tyrosine phosphorylation was carried out using lysates from cells plated onto BSA control (Con) substrate or 10 μg/ml immobilised fibronectin (Fn), laminin (Lam), fibrillin-1 (PF8) (Fib-1), collagen type I (Col I) or collagen type IV (Col IV) in serum-free conditions for 90 minutes at 37°C. PDGFR-β was isolated from equivalent MSC lysates by IP analysis using anti-PDGFR-β with anti-rabbit IgG antibodies as a control, then tyrosine phosphorylation was detected by immunoblotting (IB) using an antibody against phosphorylated tyrosine (Tyr-P) (supplementary material Table S1). Membranes were re-probed with anti-PDGFR-β antibody, as a loading control. Quantitative analysis was evaluated by densitometry with Tyr-P normalised to total PDGFR-β and is represented as the fold increase above that with BSA control substrate. A representative example of two independent experiments is shown. ECM-induced PDGFR-β (C) Y751 and (D) Y1021 PDGFR-β phosphorylation was determined using ELISAs for phosphorylated PDGFR-β, with all conditions normalised to the total amount of PDGFR-β. MSCs were plated onto 10 μg/ml immobilised ECM proteins in serum-free conditions for 90 minutes at 37°C. PDGFR-β phosphorylation is represented as the fold increase above the control BSA-induced PDGFR-β tyrosine phosphorylation, against which ECM proteins were compared (***, P<0.001 by one-way ANOVA). Experiments were performed in triplicate, on the same microtitre plate, and at least three times. Data are means±s.d. for at least three independent experiments.
ECM-induced increases in RTK phosphorylation was not due to differences in cell attachment. ECM-induced PDGFR-β tyrosine phosphorylation in serum-free conditions was also examined by immunoprecipitation and immunoblotting, normalised to the total amount of PDGFR-β (Fig. 1B). Fibroenectin induced the greatest PDGFR-β tyrosine phosphorylation activity, but cells cultured on fibrillin-1 PF8 also exhibited an increase in the level of phosphorylated PDGFR-β. By contrast, MSC adhesion to laminin or collagen types I or IV had little effect on PDGFR-β activity compared with that in cells on the BSA control substrate.

To confirm that adhesion to ECM in serum-free conditions differentially activates PDGFR-β, the phosphorylation status of specific PDGFR-β tyrosine residues was examined using immunoblot assays. Analysis of PDGFR-β tyrosine phosphorylation revealed that plating MSCs onto fibronectin, fibrillin-1 PF8 and collagen types I or IV for 90 minutes all triggered phosphorylation of Y751 (Fig. 1C) and Y1021 (Fig. 1D) above that in the basal BSA control. MSCs plated onto fibronectin had the greatest level of PDGFR-β Y751 and Y1021 phosphorylation. By contrast, laminin had no more of an effect on PDGFR-β Y751 and Y1021 phosphorylation than exposure to BSA. Because PDGFR-β phosphorylation was ECM-dependent, we checked whether the abundance of PDGFR-β, at protein and mRNA levels, was altered following the adhesion of MSCs to ECM ligands for 90 minutes in serum-free conditions. Immunoblotting, reverse-transcription-PCR (RT-PCR) and quantitative real-time PCR (qPCR) revealed that PDGFR-β expression was not increased by adhesion to the ECM over this timeframe (supplementary material Fig. S1B). RT-PCR and qPCR assays also revealed only trace levels of endogenous PDGF-BB, PDGF-DD and VEGF-A transcripts, which were not upregulated by plating MSCs onto fibroenectin for 90 minutes (supplementary material Fig. S1C). Enzyme-linked immunosorbent assays (ELISAs) showed that adhesion of MSCs to fibroenectin or laminin did not induce secretion of PDGF-BB or VEGF-A above that on the BSA control substrate (supplementary material Fig. S1D). In addition, treatment of MSCs with a PDGFR-β-neutralising antibody (AF385), which prevents PDGF-BB from binding and activating the receptor, failed to block fibroenectin-induced Y751 or Y1021 PDGFR-β activation (see Fig. 5A).

α5β1-integrin mediates fibroenectin-induced PDGFR-β tyrosine phosphorylation
Integrins are the principal receptors for ECM ligands, serving as transmembrane links between extracellular adhesion molecules and the intracellular actin cytoskeleton (Askar et al., 2009). As the integrin expression profile of a cell will dictate its interactions with the ECM, the expression of integrins by cultured MSCs was analysed by flow cytometry (supplementary material Fig. S2A). Integrin subunits α5, α3, α5 and β1, and the αβ5-integrin, were all prominently expressed by MSCs, whereas the αβ3-integrin, and integrin subunits α1, α2, α4, α6 and β4, had a lower level of expression. In comparison with the isotype-matched IgG controls, αβ1-integrin and αβ6-integrin were not detectable.

Having identified the integrins expressed by MSCs, we investigated which of these receptors mediated adhesion to two contrasting ECM proteins: fibroenectin, which induced the greatest level of PDGFR-β phosphorylation in MSCs, and laminin, which induced no substantial PDGFR-β phosphorylation (see Fig. 1). Adhesion assays were performed for 90 minutes in the presence of function-blocking monoclonal antibodies (mAbs) against integrin subunits and dimeric integrin receptors. MSC adhesion to fibroenectin was greatly reduced by the function-blocking anti-β1-integrin antibody (mAb13) and was also substantially inhibited by each of three different function-blocking antibodies against the integrin subunit α5 (mAb16, JB55 and P1D6) (supplementary material Fig. S2B). By contrast, non-inhibitory antibodies against the integrin subunits β1 (8E3) and α5 (mAb11) had no effect on attachment. MSC adhesion to fibroenectin was partially reduced by integrin function-blocking antibodies against the integrin subunits α4 (HP2/1) or αv (17E6), and the αvβ3-integrin (LM609 and 23C6). As expected, MSC adhesion to laminin was unaffected by function-blocking antibodies against the integrin subunits α5 or αv, and the αvβ3-integrin (data not shown); however, MSC attachment to laminin was almost completely blocked by the inhibitory anti-β1-integrin antibody (mAb13) (supplementary material Fig. S2C). MSC adhesion to laminin was also substantially reduced by the inhibitory anti-α1-integrin antibody (FB12) and partially reduced by the inhibitory anti-α6-integrin antibody (GoH3), but the non-inhibitory anti-β1-integrin antibody (8E3) had little effect.

To investigate whether α5β1- and αvβ3-integrin, which supported MSC adhesion to fibroenectin, modulated fibroenectin-induced PDGFR-β activity, PDGFR-β Y751 and Y1021 phosphorylation levels were assayed in the presence of function-blocking antibodies against the integrin subunits α5 and β1, and the αvβ3-integrin, or with the cyclic RGD peptide cilengitide that inhibits αv integrins (Fig. 2A). MSCs plated onto fibronectin for 90 minutes in the presence of function-blocking antibodies against the α5 (JB55) or β1 (mAb13) integrin subunits exhibited a substantial reduction in the level of PDGFR-β Y751 and Y1021 phosphorylation compared with the level of fibroenectin-induced phosphorylation (set at 100%). By contrast, the presence of inhibitory antibodies against αvβ3-integrin (23C6), cilengitide, non-functional anti-α5-integrin antibody (mAb11), or non-inhibitory anti-β1-integrin antibody (8E3), resulted in no substantial change in the level of PDGFR-β phosphorylation. As the same concentration of an anti-αvβ3-integrin antibody (23C6) was shown to reduce MSC adhesion to fibroenectin (see supplementary material Fig. S2B), failure to inhibit PDGFR-β phosphorylation was not due to the concentration of antibody used. MSCs plated onto laminin for 90 minutes had no detectable change in the level of PDGFR-β Y751 or Y1021 phosphorylation compared with that of cells on the control BSA substrate, in either the absence or presence of blocking antibodies against the integrin α5 or β1 integrin subunits or the αvβ3-integrin (data not shown). Thus, fibroenectin-induced phosphorylation of PDGFR-β in MSCs is dependent upon the α5 or β1 integrin subunits, confirming the fibroenectin receptor α5β1-integrin, but not αvβ3-integrin, as the mediator of fibroenectin-induced ligand-independent PDGFR-β phosphorylation.

To determine whether α5β1-integrin-dependent fibroenectin-induced phosphorylation of PDGFR-β is a generalised mechanism, the effect of integrins in modulating fibroenectin-induced PDGFR-β activity was examined in human aortic SMCs (Fig. 2B). In a manner similar to MSCs, SMCs plated onto fibroenectin for 90 minutes had a substantial increase in PDGFR-β Y751 and Y1021 phosphorylation compared with cells on BSA control substrate. Furthermore, SMCs on fibroenectin in the presence of function-blocking antibodies against the α5 (mAb16) or β1 (mAb13) integrin subunits exhibited a substantial reduction in the level of PDGFR-β Y751 and Y1021 phosphorylation compared with the fibroenectin-
induced phosphorylation (set at 100%). However, in contrast with MSCs, fibronectin-induced PDGFR-β activity in SMCs was also reduced in the presence of an inhibitory antibody against the αvβ3-integrin (23C6). Thus, fibronectin-induced phosphorylation of PDGFR-β in SMCs is mediated by α5β1- and αvβ3-integrin.

As fibronectin-induced PDGFR-β phosphorylation was differentially mediated by integrins in different cell types, the abundance of the integrin subunits α5 and αv at the protein level was determined following the adhesion of MSCs or SMCs to fibronectin for 90 minutes in serum-free conditions (Fig. 2C). Immunoblotting revealed that integrin α5 protein expression was similar in MSCs and SMCs, whereas integrin αv was more abundantly expressed in SMCs compared with its expression in MSCs.

**Fig. 2. Fibronectin-induced integrin-mediated PDGFR-β tyrosine phosphorylation.** The effects of inhibitory anti-integrin antibodies on fibronectin-induced PDGFR-β (i) Y751 and (ii) Y1021 phosphorylation were determined in (A) MSCs or (B) SMCs, using ELISAs for phosphorylated PDGFR-β, with all conditions normalised to the total amount of PDGFR-β. Cells in serum-free conditions were plated onto 10 μg/ml immobilised fibronectin for 90 minutes in the presence of 10 μg/ml anti-integrin inhibitory mAbs or 10 μM integrin-αv-inhibiting peptide cilengitide. Antibodies were specific for the integrin α5 subunit (JBS5, mAb11), the integrin β1 subunit (mAb13, 8E3) and αvβ3-integrin (23C6). Antibody specificity is indicated in brackets next to the antibody name; NF denotes a non-functional antibody, as a control. All conditions are expressed relative to fibronectin-induced PDGFR-β (i) Y751 and (ii) Y1021 phosphorylation (100%) in the absence of antibody (**P<0.01, ***P<0.001 by one-way ANOVA). Control (Con; broken line) indicates the basal tyrosine phosphorylation of PDGFR-β induced by BSA. All experiments were performed in triplicate, on the same microtitre plate, and at least three times. Data are means±s.d. for at least three independent experiments. (C) Expression of the integrin α5 and integrin αv subunits was examined in MSCs and SMCs plated onto 10 μg/ml immobilised fibronectin, in serum-free conditions, for 90 minutes at 37°C. (i) Protein expression was detected by immunoblotting (IB) equal amounts (10 μg) of cell lysates using anti-integrin-α5 or anti-integrin-αv antibodies. Membranes were reprobed with anti-β-actin antibody, as a loading control. A representative example of two independent experiments is shown. (ii) Quantitative analysis was evaluated by densitometry with data normalised to the level of β-actin. Data are represented as the mean pixel density (±s.d.) for two independent experiments (**P<0.01 by one-way ANOVA).

**Fibronectin induces the association of PDGFR-β and α5β1-integrin in serum-free conditions**

Crosstalk mechanisms between PDGFR-β and α5β1-integrin in MSCs might involve physical interactions between the integrin and PDGFR-β receptors and/or intracellular signalling events leading to PDGFR-β activation. Co-immunoprecipitation experiments were conducted in serum-free conditions to examine receptor associations in MSCs plated onto fibronectin or laminin for 90 minutes (Fig. 3A). Co-immunoprecipitation of PDGFR-β and integrin subunit α5 occurred in lysates derived from MSCs plated onto fibronectin. By contrast, no association of PDGFR-β and integrin subunit α5 was detected in lysates derived from SMCs plated onto laminin.
To determine whether the stimulatory effect of fibronectin on PDGFR-β activation was attributable to a physical interaction between the extracellular regions of PDGFR-β and α5β1-integrin, surface plasmon resonance (BIAcore) analysis was performed (supplementary material Fig. S3A,B). No direct interaction was detected between the extracellular domains of either PDGFR-β and α5β1-integrin, surface plasmon resonance (BIAcore) analysis was performed (supplementary material Fig. S3A,B). No direct interaction was detected between the extracellular regions of PDGFR-β and α5β1-integrin, and fibronectin (50 kDa adhesion fragment) bound PDGFR-β and α5β1-integrin, respectively. However, both the extracellular domains of α5β1-integrin and PDGFR-β (K<sub>d</sub>=19 nM) strongly bound immobilised heparin (supplementary material Fig. S3C), and fibronectin-induced co-immunoprecipitation of PDGFR-β and integrin subunit α5 was prevented by excess heparin and treatment with K5 heparan lyase (supplementary material Fig. S3D). Thus, cell surface heparan sulphate proteoglycans mediate the association between PDGFR-β and α5β1-integrin.

**Fibronectin induces temporal phosphorylation of PDGFR-β and colocalisation with α5β1-integrin in serum-free conditions**

Maximal fibronectin-induced Y751 and Y1021 PDGFR-β phosphorylation was detected at 90 minutes by immunoblotting, compared with that upon exposure for 15 or 240 minutes (Fig. 3B). Temporal phosphorylation of PDGFR-β was confirmed by phosphorylated RTK array analysis (supplementary material Fig. S4A). Fibronectin also induced tyrosine phosphorylation of other RTKs throughout the timecourse of 15, 90 and 240 minutes, notably PDGFR-α and EGFR, which were induced in a fibronectin-dependent manner (see Fig. 1A), but the phosphorylation remained constant over the time period examined (supplementary material Fig. S4A). Although the level of PDGFR-α phosphorylation was of lower magnitude compared with that of PDGFR-β phosphorylation under the same conditions, an ELISA for phosphorylated PDGFR-α showed that MSCs plated onto fibronectin for 90 minutes in the presence of function-blocking antibodies against integrin subunits α5 (JBS5), β1 (mAb13) or αv (17E6), or cilegitudine, exhibited a substantial reduction in the level of PDGFR-α Y742 phosphorylation compared with the fibronectin-induced phosphorylation (set at 100%) (supplementary material Fig. S4B). Thus, whereas fibronectin-α5β1-integrin engagement mediates PDGFR-β activity in MSCs, fibronectin-induced phosphorylation of PDGFR-α is mediated by α5β1-integrin and αv integrins.

To investigate further the relationship between PDGFR-β and α5β1-integrin in MSCs following adhesion to fibronectin, the cellular distribution of phosphorylated PDGFR-β and integrin subunit α5 was examined by immunofluorescence microscopy. PDGFR-β phosphorylated on Y751 and Y1021 was detected at the cell periphery following a 15-minute incubation on fibronectin (Fig. 4A,Bi), whereas MSCs had not adhered to laminin at this timepoint (data not shown). After MSCs had been plated for 90 minutes on fibronectin (Fig. 4A,Bii), but not laminin (supplementary material Fig. S5A), PDGFR-β Y751 and Y1021 phosphorylation was detected at the leading edge and in a tidemark of integrin-α5-containing focal adhesions behind the leading edge.

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Fig. 3. Fibronectin-induced association of PDGFR-β and α5β1-integrin. (A) The association of PDGFR-β with integrin subunit α5 was examined by immunoprecipitation (IP). MSCs in serum-free conditions were incubated on tissue culture plastic, as a control (Con), or on 10 μg/ml immobilised fibronectin (Fn) or laminin (Lam), for 90 minutes at 37°C. (i) PDGFR-β was isolated from equivalent MSC lysates by IP using anti-PDGFR-β antibody, with anti-rabbit IgG antibody as a control, then integrin subunit α5 association was detected by immunoblotting (IB) using an antibody against integrin α5. Membranes were re-probed with anti-PDGFR-β as a loading control. (ii) Integrin subunit α5 was isolated by IP using antibodies against integrin α5 or IgG, as a control, then PDGFR-β association was detected by IB analysis using anti-PDGFR-β antibody. Membranes were reprobed with an antibody against integrin α5 as a loading control. A representative example of three independent experiments is shown. (B) Immunoblotting of PDGFR-β Y751 or Y1021 phosphorylation was carried out using MSC lysates, taken at 15, 90 or 240 minutes, from wells coated with 10 μg/ml fibronectin. (i) Levels of PDGFR-β phosphorylation were detected in equal (10 μg) amounts of cell lysates using antibodies against phosphorylated PDGFR-β Y751 or Y1021, or anti-PDGFR-β antibody, as a loading control. A representative example of two independent experiments is shown. (ii) Quantitative analysis, evaluated by densitometry, with data normalised to the total amount of PDGFR-β. Data are means±s.d. for two independent experiments (**P<0.001 compared with other time points).
Fibronectin-induced colocalisation of phosphorylated PDGFR-β and integrin subunit α5 at 90 minutes was supported by colocalisation analysis (Fig. 4C). After 4 hours of incubation on fibronectin (Fig. 4A,Bii), PDGFR-β phosphorylated on Y751 and Y1021 was detected predominantly at the leading edge of MSCs, whereas integrin subunit α5 was present towards their trailing edge. Thus, in serum-free conditions, fibronectin first induces PDGFR-β phosphorylation at the leading edge, followed by transient colocalisation of phosphorylated PDGFR-β with α5β1-integrin in focal adhesions, then dissociation and translocation of α5β1-integrin towards the rear of the cells.

Fibronectin and PDGF-BB additively contribute to PDGFR-β tyrosine phosphorylation

Given that MSC adhesion to fibronectin in serum-free conditions was shown to induce ligand-independent phosphorylation of PDGFR-β and association with α5β1-integrin, we investigated the effect of fibronectin on PDGF-BB-stimulated PDGFR-β phosphorylation. PDGF-BB, the major ligand of PDGFR-β, is an important mitogen and chemoattractant for mesenchymal cells (Betsholtz et al., 2001). ELISA analysis demonstrated that, compared with ligand-independent fibronectin-induced PDGFR-β activity, exposure to 50 ng/ml PDGF-BB increased the levels of...
Y751 (Fig. 5Ai) and Y1021 (Fig. 5Aii) phosphorylation by 78% and 83%, respectively. The addition of PDGF-AA, which binds only the PDGF-α homodimer, was used as a negative control and did not induce PDGF-β activity above that induced by fibronectin (data not shown). MSCs cultured on laminin did not induce receptor phosphorylation above that with BSA, although PDGF-BB did induce a substantial increase in PDGF-β activity (Fig. 5A). These results show that fibronectin and PDGF-BB both contribute to PDGF-β phosphorylation, whereas on laminin PDGF-β phosphorylation is only induced in the presence of PDGF-BB.

To confirm that the fibronectin-enhanced PDGF-BB stimulation of PDGF-β Y751 and Y1021 phosphorylation in MSCs was α5β1-integrin-dependent, the effects of the function-blocking antibody against integrin subunit α5 (JBS5) and/or a neutralising anti-PDGF-β (AF385) antibody, which inhibits receptor phosphorylation, were examined (Fig. 5A). In the absence of the PDGF-BB ligand, only JBS5 substantially reduced the level of PDGF-β phosphorylation, to near the basal levels induced by BSA, whereas AF385 had no detectable effect. However, when MSCs plated onto fibronectin were stimulated with PDGF-BB, addition of either JBS5 or AF385 resulted in a substantial reduction in PDGF-BB Y751 and Y1021 phosphorylation levels, compared with the levels in MSCs in the absence of inhibitory antibodies. Phosphorylation of Y751 and Y1021 was further reduced when JBS5 and AF385 were plated onto 10 μg/ml immobilised fibrillin-1 (PF8) in the presence of PDGF-BB (**P<0.001 by one-way ANOVA). Antibody specificity is indicated in brackets next to the antibody name; NF denotes a non-functional antibody, as a control. All conditions are expressed relative to PF8-induced PDGF-BB (**P<0.001 by one-way ANOVA). Control (Con; broken line) indicates the basal tyrosine phosphorylation of PDGFR-β (100%) in the absence of antibody blocking experiments show that fibronectin-dependent regulation of PDGF-β enhances PDGF-BB-stimulated PDGF-β Y751 and Y1021 phosphorylation. We also found that PDGF-BB directly binds α5β1-integrin (supplementary material Fig. S3B), suggesting that receptor co-clusters of α5β1-integrin with PDGF-β might enhance the interaction of PDGF-BB with its receptor. However, although PDGF-β was also shown to be activated by the α5β1-integrin-binding fragment (PF8) of fibrillin-
1 (see Fig. 1) in an α5β1-integrin-dependent (but not integrin αvβ3-dependent) manner (Fig. 5B), in contrast with fibronectin-enhanced PDGF-BB stimulation of PDGFR-β phosphorylation. PDGF-BB had no further effect on PDGFR-β Y751 and Y1021 phosphorylation when MSCs were cultured on PF8 (Fig. 5B).

**PDGFR-β and α5β1-integrin crosstalk mediates MSC migration**

We next investigated the functional importance of the crosstalk between α5β1-integrin and PDGFR-β for MSC migration. In serum-free conditions, MSC migration in fibronectin-coated Boyden chambers was increased by ~2.8-fold above that on BSA-coated inserts, whereas laminin had no effect above that with the basal BSA control (Fig. 6A). PDGF-BB substantially increased MSC migration towards fibronectin, laminin and BSA above that induced by serum-free conditions (Fig. 6A). To confirm that the fibronectin-enhanced PDGF-BB stimulation of MSC migration was α5β1-integrin-dependent, the effects of JBS5 and/or AF385 antibody on MSC migration were examined (Fig. 6B). In the absence of PDGF-BB ligand, only JBS5 inhibited MSC migration, whereas the anti-PDGFR-β antibody had no detectable effect. MSCs exposed to PDGF-BB in the presence of JBS5 or AF385 resulted in a substantial reduction in MSC migration towards fibronectin, compared with the amount of migration in the absence of inhibitory antibodies. MSC migration towards fibronectin was further reduced upon exposure to PDGF-BB in the presence of both JBS5 and AF385 (*ΔΔΔ*P<0.001 by one-way ANOVA). All data are the mean (±s.d.) optical density (OD 570 nm) of stained migratory cells from two readings in an individual experiment, repeated three times. Images above each bar graph are representative of the migratory cells per field on the membrane underside of three independent experiments. Scale bar: 100 μm.

**Fibronectin–α5β1-integrin engagement and FAK activation mediates PDGFR-β-induced PI3K activity and actin arrangement**

Having established that fibronectin- and α5β1-integrin-dependent regulation of PDGFR-β enhances PDGF-BB-stimulated PDGFR-β phosphorylation and MSC migration, we examined the...
involvement of specific intracellular signalling pathways. FAK is an important receptor-proximal link between PDGFR and integrin signalling pathways (Sieg et al., 2000). Therefore to determine whether FAK integrates the signalling pathways activated by both fibronectin-α5β1-integrin engagement and PDGFR-β, FAK activation at the major autophosphorylation Y397 site was examined by immunoblotting (Fig. 7Ai). In a manner similar to ECM-induced PDGFR-β phosphorylation, MSCs plated onto fibronectin for 90 minutes induced an increase in FAK Y397 phosphorylation, whereas MSCs plated onto laminin or BSA had no detectable FAK Y397 phosphorylation. MSCs stimulated with PDGF-BB showed markedly increased fibronectin-induced phosphorylation of FAK. By contrast, however, MSCs exposed to PDGF-BB when plated on laminin did not display an increase in FAK Y397 phosphorylation, suggesting that MSC adhesion to fibronectin, and engagement of α5β1-integrin, is necessary for PDGFR-BB-induced FAK phosphorylation. To confirm that fibronectin-induced PDGFR-BB-stimulated phosphorylation of FAK was α5β1-integrin-dependent, the effect of JBS5 was examined (Fig. 7Ai). In the absence or presence of PDGF-BB ligand, FAK phosphorylation was markedly reduced by adding JBS5. Furthermore, partial knockdown of FAK by small interfering RNA (siRNA) was sufficient to substantially reduce fibronectin-induced PDGFR-β Y751 and Y1021 phosphorylation levels determined using an ELISA for phosphorylated PDGFR-β, with all conditions normalised to that with total PDGFR-β. MSCs transfected with 3 μg of siRNA FAK or scrambled siRNA, as a control, were plated onto 10 μg/ml immobilised fibronectin for 90 minutes at 37°C. Conditions are expressed relative to fibronectin-induced PDGFR-β (Y397), (ii) phosphorylated Akt (S473) or (iii) phosphorylated PLCγ-1 normalised to the level of β-actin. Data are the fold increase above that with basal control substrate. A representative example of two independent experiments is shown. (B) The effect of FAK siRNA knockdown on fibronectin-induced PDGFR-β Y751 and Y1021 phosphorylation levels was determined using an ELISA for phosphorylated PDGFR-β, with all conditions normalised to that with total PDGFR-β. MSCs transfected with 3 μg of siRNA FAK or scrambled siRNA, as a control, were plated onto 10 μg/ml immobilised fibronectin for 90 minutes at 37°C. Conditions are expressed relative to fibronectin-induced PDGFR-β Y751 or Y1021 phosphorylation (100%) in the presence of scrambled siRNA (**P<0.001 by one-way ANOVA). Experiments were performed in triplicate, on the same microtitre plate, and at least three times. Data are the means±s.d. for at least three independent experiments. The knockdown efficiency of the FAK siRNA as assessed by immunoblotting is shown below the graph. (C) The effect of FAK siRNA knockdown on fibronectin-induced Akt S473 phosphorylation levels was determined by immunoblotting. MSCs transfected with 3 μg of siRNA FAK or scrambled siRNA, as a control, were plated onto 10 μg/ml immobilised fibronectin for 90 minutes at 37°C. (i) Phosphorylation levels of Akt were detected in equal (10 μg) amounts of cell lysates using an antibody against phosphorylated Akt S473 (S473). The membrane was re-probed with anti-β-actin antibody as a loading control. A representative example of two independent experiments is shown. (ii) Quantitative analysis, as determined by densitometry, with the levels of phosphorylated Akt normalised to those of β-actin. Data are the means±s.d. for two independent experiments (**P<0.01 by Student’s t-test). The knockdown efficiency of the FAK siRNA is shown in B.
MSCs plated onto fibronectin for 90 minutes induced S473 phosphorylation of Akt (Fig. 7Aii) and Y783 phosphorylation of PLCγ-1 (Fig. 7Aiii). By contrast, MSCs plated onto laminin or BSA, displayed neither Akt nor PLCγ-1 phosphorylation. MSC stimulation with PDGF-BB markedly increased fibronectin-induced phosphorylation of both Akt and PLCγ-1. Fibronectin-induced S473 Akt phosphorylation in the absence or presence of PDGF-BB, was abolished by adding JBS5 (Fig. 7Aii), indicating that α5β1-integrin regulates PDGFR-β-ligand-induced PI3K activity in MSCs. By contrast, incubation of MSCs with JBS5
did not alter PLCγ-1 phosphorylation (Fig. 7Aiii). Furthermore, partial knockdown of FAK by siRNA was sufficient to substantially reduce fibronectin-induced Akt S473 phosphorylation levels (Fig. 7C). Thus, FAK is a mediator of fibronectin- and α5β1-integrin-dependent regulation of Akt-P13K activity.

PDGF-BB induces cell migration in conjunction with marked reorganisation of actin filaments and the appearance of F-actin membrane ruffles (Meima et al., 2009), whereas mutant PDGF-stimulated cells expressing the non-phosphorylatable Tyr-to-Phe (Y751F) PDGF-BB lack ruffled edges (Wenström et al., 1994). Our finding that α5β1-integrin engagement of fibronectin regulates PDGF-BB-mediated Akt-P13K activity in MSCs led us to examine the effect of α5β1-integrin inhibition on F-actin organisation and the localisation of Y751 phosphorylated PDGF-BB (Fig. 8A). Plating MSCs on fibronectin in serum-free conditions induced Y751 phosphorylated PDGF-BB clustering and membrane ruffling; these cellular changes were more pronounced in the presence of PDGF-BB, with intensely stained patches of actin at the rim of the cells. Inhibition of α5β1-integrin using the JBS5 function-blocking antibody substantially reduced the clustering of fibronectin-induced Y751-phosphorylated PDGF-BB and F-actin in membrane ruffles, both in the absence and presence of PDGF-BB (Fig. 8A). By contrast, when MSCs were plated onto laminin, Y751-phosphorylated PDGF-BB and F-actin clustering were only induced in the presence of PDGF-BB (supplementary material Fig. S5C).

As α5β1-integrin engagement of fibronectin regulated PDGF-BB-mediated Akt-P13K activity (Fig. 7Aii) and F-actin organisation (Fig. 8A), we investigated the effect of P13K inhibition using the chemical inhibitor LY294002 on MSC F-actin organisation and migration. Inhibition of P13K activity ablated both fibronectin- and PDGF-BB-induced F-actin membrane ruffles (Fig. 8B), and substantially attenuated MSC migration towards fibronectin in the absence or presence of PDGF-BB (Fig. 8C).

In summary, our results show that the adhesion of MSCs to fibronectin through α5β1-integrin both directly stimulates PDGF-BB and potentiates PDGF-BB-stimulated PDGF-BB signalling. In turn, PDGF-BB-mediated Akt-P13K activation drives the cytoskeletal changes that regulate cell migration.

**Discussion**

The crucial importance of PDGF-BB in directing vascular cell behaviour is well documented, yet ECM-dependent mechanisms that regulate its signalling are not well understood. Using human MSCs, we have shown that adhesion to fibronectin specifically induces PDGF-BB signalling in an α5β1-integrin-dependent manner (Fig. 9). Fibronectin regulates PDGF-BB-dependent actin reorganisation and cell migration in the absence of growth factor ligand and also potentiates PDGF-BB-induced PDGF-BB signalling. Thus, fibronectin-rich matrices are crucial regulators of PDGF-BB-mediated mesenchymal cell migration during vascular remodelling.

Nascent blood vessels contain many ECM molecules, including fibronectin, laminin, collagens and fibrillins. Our study shows that PDGF-BB signalling is differentially affected by cell adhesion to specific ECM ligands. In particular, adhesion to fibronectin and fibrillin-1, but not laminin, induces tyrosine phosphorylation of PDGF-BB in a manner independent of growth factor ligand. The stimulatory effect of fibronectin (a key regulator of cell adhesion, migration and survival during tissue formation, and a regulator of wound repair) is integrin-dependent as blocking α5β1-integrin but not αvβ3-integrin prevented PDGF-BB activation. Thus ECM, through specific integrins, directly stimulates PDGF-BB.

PDGF-BB activation induces dynamic changes in the cytoskeleton and membrane ruffles during cell migration (Mellström et al., 1988; Ruusala et al., 2008). We found that, in MSCs on fibronectin, crosstalk with α5β1-integrin regulates PDGF-BB-mediated cell adhesion, spreading and migration. Adhesion to fibronectin induces a rapid movement of phosphorylated PDGF-BB to the leading edge of MSCs, and a tidemark of phosphorylated PDGF-BB transiently colocalises with α5β1-integrin-containing focal adhesions behind the leading edge. Thus, PDGF-BB activity is a crucial element of α5β1-integrin-mediated adhesion and migration in these cells. Although co-immunoprecipitations showed that PDGF-BB and integrin subunit α5 associate in complexes, even in the absence of PDGF, we did not detect a direct interaction between the extracellular domains of PDGF-BB and α5β1-integrin. However, we did find that both receptor ectodomains strongly bound heparin, and fibronectin-induced co-immunoprecipitation of PDGF-BB and integrin subunit α5 was prevented by excess heparin and treatment with K5 heparan lyase. Thus, cell surface heparan sulphate proteoglycans mediate the association between PDGF-BB and α5β1-integrin. The receptor crosstalk could also involve molecules such as pericellular tissue transglutaminase, which binds to PDGF-BB (Zemskov et al., 2009), or tetraspanins, which form membrane complexes with integrin receptors and are implicated in integrin-mediated cell migration (Berditchevski and Odintsova, 1999).

Previous studies have indicated that the β1 integrin subunit and αvβ3-integrin can influence PDGF-BB activity (Sundberg and
Rubin, 1996; Schneller et al., 1997; Woodard et al., 1998; Borges et al., 2000; Minami et al., 2007; Amano et al., 2008; Zemskov et al., 2009). Our study is the first to identify the major fibronectin α5β1-integrin as a key partner in PDGFR-β-mediated mesenchymal cell migration, both in the absence of growth factors and in potentiating PDGF-BB-mediated MSC migration. Integrin-mediated activation of PDGFR-β could occur as a consequence of integrin clustering, which is known to alter the spatial distribution of PDGFR-β within focal adhesions (Burridge et al., 1988; Miyamoto et al., 1996; Boudreau and Jones, 1999) enhancing PDGFR-β dimerisation and activation. This hypothesis is supported by our finding that α5β1-integrin engagement of fibronectin and PDGF-BB can induce both clusters of phosphorylated PDGFR-β and membrane ruffles, and that PDGF-BB can bind α5β1-integrin, which might further enhance the activity of PDGFR-β. Interactions of phosphorylated PDGFR-β with cytoskeletal proteins have been reported (Schneller, 2001), but it is not known whether PDGFR-β can directly bind actin. However, EGFR binds actin (den Hartigh et al., 1992; Tang and Gross, 2003) and localises in signalling complexes at membrane ruffles (Diakonova et al., 1995), thereby enhancing signalling efficiency (Gronowski and Bertics, 1993).

We have found that crosstalk between PDGFR-β and α5β1-integrin induce synergistic signalling responses. FAK and Akt-PISK activity was cooperatively stimulated by fibronectin and PDGF-BB in an α5β1-integrin-dependent manner, whereas FAK knockdown reduced fibronectin-induced phosphorylation of PDGFR-β and Akt phosphorylation, thereby linking FAK and Akt-PISK signals. This is in agreement with a previous study that found FAK to be an important receptor-proximal link between PDGFR and integrin signalling pathways (Siegl et al., 2000). Inhibition of Akt-PISK inhibited both fibronectin- and PDGF-induced actin reorganisation and migration of MSCs. These findings are consistent with previous reports that show a PDGFR-β mutant that is unable to bind and activate PISK fails to mediate actin reorganisation and chemotaxis (Wennström et al., 1994). Moreover, in the absence of PDGFR-β-driven PISK signalling, epicardial cells adopt an irregular actin cytoskeleton, leading to aberrant migration into the myocardium and defective coronary artery formation (Mellgren et al., 2008).

Thus, ECM controls mesenchymal cells through crosstalk with the potent vascular receptor PDGFR-β.

**Materials and Methods**

**Cells and reagents**

Human MSCs derived from the normal bone marrow of five different individuals (CD44, CD73 and CD105 positive and CD11b, CD14, CD34 and CD45 negative; from 28- and 34-year-old females and 19-, 25- and 33-year-old males; Lonza) were tested for their ability to differentiate into osteogenic, adipogenic and chondrogenic lineages. MSCs were maintained as previously described (Ball et al., 2007). Human aortic SMCs (32-year-old female; Lonza) were maintained in SMC growth medium (Invitrogen). Cells were cultured on human plasma fibronectin (Chemicon), murine laminin (α1β1γ1), bovine collagen type I, murine collagen type IV (BD Biosciences), recombinant fibronectin-1 protein fragment (PF8) (prepared in-house) (Cain et al., 2005) or a recombinant 50-kDa fibronectin fragment comprising type III repeats 6–10 (a gift from Martin Humphries, University of Manchester, Manchester, UK). All growth factors and the anti-PDGFR-β neutralising antibody (AF385) were obtained from R&D Systems. The cyclic RGD-blocked peptide cilengitide (Nisato et al., 2003) was a gift from Simon L. Goodman, Merck, Garching, Germany, and the P13K inhibitor LY294002 was from Merck. For the antibodies used, see supplementary material Table S1.

**Phosphorylated RTK array**

A human phosphorylated RTK array kit (R&D Systems) was used to simultaneously detect the relative tyrosine phosphorylation levels of 42 different RTKs, as previously described (Ball et al., 2007).

**Immunoblotting**

Isolated proteins were resolved using pre-cast Tris-acetate (3–8%) or Bis-Tris (4–12%) gels (Invitrogen), transferred onto nitrocellulose, incubated with primary antibody overnight and then incubated with horseradish peroxidase (HRP)-conjugated secondary antibody for 2 hours. Detection of proteins was performed with enhanced chemiluminescence (ECL) western blotting reagent, as previously described (Ball et al., 2007). The pixel density of bands was determined with the Gene Tools v3 software.

**Immunoprecipitation**

Cell lysates (500 µg) were pre-cleared using 10% protein-A–Sepharose, immunoprecipitated with primary antibody overnight, and then the immunocomplexes were isolated by incubation with 10% protein-A–Sepharose for 2 hours, as previously described (Ball et al., 2007).

**siRNA transfection**

MSCs (5×10⁵ cells), together with 3 µg of small interfering RNA (siRNA), were transfected by electroporation using a human Amaxa Nucleofector kit (Lonza), then cultured in growth medium overnight. Validated siRNA, functionally tested to provide ≥70% gene knockdown for FAK, was obtained from Qiagen (S02622130), and scrambled siRNA was used as a control (Qiagen).

**Phosphorylated PDGFR immunoblot**

Phosphorylated PDGFR-β immunoblot was used to measure the levels of tyrosine residue (Y751 and Y1021) phosphorylation for PDGFR-β, according to the manufacturer’s instructions. Assays were modified to measure phosphorylated PDGFR-α (Y742) utilising an antibody against phosphorylated PDGFR-α (Y742) (R&D Systems).

**Immunofluorescence analysis**

Cells were fixed using 4% paraformaldehyde for 20 minutes, permeabilised with 0.5% Triton X-100 for 4 minutes, blocked with 3% fish-skin gelatin (Sigma–Aldrich) for 1 hour and then incubated with primary antibody overnight at 4°C. Alexa-Fluor-488 or -594-conjugated secondary antibodies (Invitrogen), and Rhodamine-conjugated phalloidin (Invitrogen) to stain filamentous actin, were added for 2 hours, then coverslips were mounted using Vectashield containing DAPI (Vector Laboratories). Images were collected on a wide-field microscope (Leica DM RXA) using a 60× oil objective and captured using a Coolsnap EZ camera (Photometrics) driven by MetaVue Software (Molecular Devices). For colocalisation analysis, images were analysed using ImageJ software and the colocalisation analysis plugin. For receptor distribution and clustering analysis, images were analysed using the ‘analyse particles’ function of ImageJ software (Chen et al., 2010). Similar best-fit lower threshold values were determined for each image to reduce signal background, with the upper threshold always set at 255. Particle sizes for colocalisation were set at a minimum of 1 pixel and maximum of 500 pixels, and colocalisation is represented by a yellow image. Particle sizes for receptor distribution and clustering were set at a minimum of 1 or 10 pixels, respectively, and a maximum of 500 pixels, and are represented by a black and white image.

**Cell migration assay**

Modified Boyden chamber assays were conducted using filter inserts of 8-µm pore size and 6.5-mm diameter (BD Biosciences), as previously described (Ball et al., 2007). Migratory cells were imaged by phase-contrast microscopy (Leica DM RXA brightfield) and captured using a Coolsnap EZ camera driven by MetaVue software.

**Statistical analysis**

In all quantification experiments, results are expressed as means±s.d. Statistical differences between two sets of data were determined using an unpaired Student’s t-test and differences between more than two data sets were determined by one-way ANOVA using the Tukey comparison test. *P<0.05 was considered statistically significant. All statistical calculations were performed using the GraphPad Prism 5 software.

This study was funded by the Medical Research Council (UK). We thank Stuart Cain for assistance with the BIACore experiments,
References


Figure S1.

A (i) Cell attachment (%)

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B (i) Cell lysate

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C (ii) RT-PCR products

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D PDGF-BB Standard curve

y = 0.0014x + 0.0034
R² = 0.9999

VEGF-A165 Standard curve

y = 0.0017x + 0.0408
R² = 0.9828

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Figure S2.

A
(i) Integrin $\alpha_1$
(ii) Integrin $\alpha_2$
(iii) Integrin $\alpha_3$
(iv) Integrin $\alpha_4$
(v) Integrin $\alpha_5$
(vi) Integrin $\alpha_6$
(vii) Integrin $\alpha_6\beta_1$
(viii) Integrin $\alpha_v$
(ix) Integrin $\beta_1$
(x) Integrin $\beta_4$
(xi) Integrin $\alpha_6\beta_3$
(xii) Integrin $\alpha_6\beta_5$
(xiii) Integrin $\alpha_6\beta_6$
(xiv) Cells only
(xv) Anti-Mouse FITC
(xvi) Anti-Rat FITC

B
Cell attachment relative to fibronectin (%)

C
Cell attachment relative to laminin (%)
Figure S3.

A) PDGFR-β immobilised on chip

B) Integrin α5β1 immobilised on chip

C) Heparin (dp20) immobilised on chip

D) IB: PDGFR-β 190-kD
   IB: β-actin 42-kD
   IP: Integrin α5
   IP: IgG
   FN Hep K5

PDGFR-β Kd = 19 nM
A (i) 15 Minutes

(ii) Pixel density (Fold increase over basal)

B

PDGFR-α Y724 phosphorylation relative to fibronectin-induced phosphorylation (%)
A

(i) Y751

merge  PDGFR-β  Integrin α5

(ii) Y1021

B

(i) Fibronectin

(ii) Fibronectin + PDGF-BB

Fibronectin

LM609

C

merge  Phalloidin  PDGFR-β Y751

Laminin

Laminin + PDGF-BB

PDGFR-β Y751 Distribution

PDGFR-β Y751 Clustering

49 ± 12

3 ± 2

98 ± 22

15 ± 7

Upper chamber

Membrane underside

Lower chamber

Fibronectin

BB

BB
Table S1. List of antibodies used in this study

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