Role of connexins in metastatic breast cancer and melanoma brain colonization

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Breast cancer and melanoma cells commonly metastasize to the brain using homing mechanisms that are poorly understood. Cancer patients with brain metastases display poor prognosis and survival due to the lack of effective therapeutics and treatment strategies. Recent work using intravital microscopy and preclinical animal models indicates that metastatic cells colonize the brain specifically in close contact with the existing brain vasculature. However, it is not known how contact with the vascular niche promotes microtumor formation. Here, we investigate the role of connexins in mediating early events in brain colonization using transparent zebrafish and chicken embryo models of brain metastasis. We provide evidence that breast cancer and melanoma cells utilize connexin gap junction proteins (Cx43, Cx26) to initiate brain metastatic lesion formation in association with the vasculature. RNAi depletion of connexins or pharmacological blocking of connexin-mediated cell-cell communication with carbenoxolone inhibited brain colonization by blocking tumor cell extravasation and blood vessel co-option. Activation of the metastatic gene twist in breast cancer cells increased Cx43 protein expression and gap junction communication leading to increased extravasation, blood vessel co-option, and brain colonization. Conversely, inhibiting twist activity reduced Cx43-mediated gap junction coupling and brain colonization. Database analyses of patient histories revealed increased expression of connexins 26 and 43 in primary melanoma and breast cancer tumors, respectively, which correlated with increased cancer recurrence and metastasis. Together our data indicate that connexins 43 and 26 mediate cancer cell metastasis to the brain and suggests that connexins might be exploited therapeutically to benefit cancer patients with metastatic disease.
**Introduction**

Approximately half of all cancer patients develop brain metastases leading to 20 percent of cancer related deaths annually (Nussbaum et al., 1996; Lu et al., 2009). Breast and skin (melanoma) cancers are among the most common cancers that metastasize to the brain with approximately 10 to 20 percent of breast cancer patients and 40 to 60 percent of melanoma patients developing brain metastases. Patients with cancer metastasis to the brain have an extremely poor prognosis with less than ten months average survival time (Nussbaum et al., 1996; Lu et al., 2009). Treatment of brain cancer metastases is impeded by low permeability of the blood brain barrier to therapeutic agents, high sensitivity of brain tissue to irradiation, and the high number of advanced metastatic lesions at the time of diagnosis (Lu et al., 2009). Therefore, new molecular targets and therapeutic strategies that aim at brain metastatic lesions are sorely needed.

Although it is not clear why breast cancer and melanoma cells show a propensity to colonize the brain, brain micrometastases are initiated along the preexisting brain vasculature in close contact with the vessel surface (Kienast et al., 2010; Lorger and Felding-Habermann, 2010; Carbonell et al., 2009, Polyak and Weinberg, 2009). This suggests that the microenvironment of the vascular niche supports cancer cell survival and propagation during the early stages of brain colonization. However, the specific mechanisms used by metastatic cells to attach to the brain vasculature and propagate are poorly understood.

Connexin-mediated gap junctions (GJs) are strongly expressed in the vessel endothelium where they maintain vessel architecture and facilitate interactions with glial cells, which surround the vessel wall (Brisset et al., 2009; Figueroa and Duling, 2009). Glial cells play a key role in the maintenance of the blood-brain barrier by facilitating strong GJ-mediated adhesive interactions with the endothelium and by regulating heterocellular ion and small molecule transport activities (Simard et al., 2003; Balabanov and Dore-Duffy, 1998; Nagasawa et al., 2006). Similarly, metastatic tumors cells, which express connexins, may adhere to and propagate along the connexin-rich vasculature by forming heterotypic GJs with the endothelium and/or glia. Heterotypic GJ formation with stromal elements could also provide the means for the exchange of life sustaining ions and immunomodulatory factors, which are necessary to establish the tumor cell-vascular niche and initiate microtumor formation. However, the role of connexins in mediating brain metastasis has not been defined and their role in local and systemic cancer cell dissemination is controversial (reviewed by Laird, 2006; Naus and Laird, 2010).

Connexins have been reported to exhibit tumor suppressor-like activities, to cause cancer cell dedifferentiation (mesenchymal to epithelial transition), and to inhibit metastasis (Laird et al., 1999; Naus and Laird, 2010; Ito et al., 2002). Also, mice with Cx43 inactivating
mutations when crossed with the ErbB2 overexpressing mice show delayed onset of palpable mammary tumors while displaying increased pulmonary metastases (Plante et al., 2011). On the other hand, in melanoma, breast, and gastric cancers connexin 43 (Cx43) and 26 (Cx26) are upregulated in invasive lesions and in cells that disseminate to the lymph nodes (Naoi et al., 2007; Haass et al., 2010; Saito-Katsuragi et al., 2007; Elzarrad et al., 2008; Chao et al., 2012). Interestingly, high expression of Cx26 was detected in invasive, perivascular areas of primary melanomas, suggesting that heterotypic GJ-mediated cell-cell adhesion and communication contributes to melanoma metastasis (Haass et al., 2010; Saito-Katsuragi et al., 2007). Finally, metastatic breast cancer cells selected for their ability to home to the brain showed increased Cx43 expression (Bos et al., 2009). These findings lead one to believe that connexins may be differentially regulated during tumor cell dissemination. It is also possible that connexin expression may be restricted to distinct and rare tumor cell types that target specific organs like the brain, which is highly enriched in connexins. In any case, the conflicting data likely reflect the differences in experimental systems, the cellular heterogeneity of tumors, compensation by other connexin family members, and the complexity of the multi-step metastatic process in general. Reconciling these critical issues is important and warrants additional investigations. In this study, we examine the role of connexins Cx43 and Cx26 in mediating metastatic breast and melanoma cell brain colonization, respectively. For these studies, we use transparent zebrafish and chicken brain metastasis models combined with high-resolution fluorescence imaging methods to monitor the early events that initiate microtumor formation at the tumor cell-vascular interface.
Results

Connexin expression in breast cancer and melanoma patients correlates with poor patient outcome. We first examined whether connexin expression in cancer patients correlates with patient prognosis and metastasis. We focused our attention on connexin isoforms 43 and 26, since they are the primary isoforms reported to be involved in breast cancer and melanoma metastasis (Elzarrad et al., 2008; Ito et al., 2002; Naoi et al., 2007; Haass et al., 2010; Saito-Katsuragi et al., 2007; Chao et al., 2012). To this end, we used the Oncomine (www.oncomine.org) cancer database that links various human gene expression profiles from primary and metastatic cancers to patient history. Although Oncomine did not specifically delineate cancer patients with brain metastases, it did reveal that increased Cx43 and Cx26 expression in primary breast cancer tumors correlated with poor patient survival and cancer recurrence (supplementary material Fig. S1A,B,H). In melanoma, increased Cx43 and 26 expression in primary lesions correlated with metastasis and poor patient survival (supplementary material Fig. S1C-G).

To determine the Cx43 expression pattern in established human brain metastases, we obtained rare brain metastases that had been surgically removed from 4 different breast cancer patients. In all four patients, Cx43 expression was significantly increased in the metastatic nodules compared to normal uninvolved brain tissue from the same patient sample (Fig. 1). However, the majority of Cx43 staining co-localized with the glial cell marker glial fibrillary acidic protein (GFAP). In fact, Cx43 immunoreactivity was not detected in the majority (>95%) of tumor cells present in the metastatic nodules, which are easily recognized by their large DAPI-stained nuclei and lack of GFAP expression (Fig. 1). Interestingly, close examination of the tumor cell-vascular interface in metastatic nodules by high-resolution imaging revealed a small population (<5%) of Cx43-positive tumor cells contacting the blood vessel surface, which did not co-localize with GFAP (Fig. 1E). Although the patient sample size is small, these findings provide initial evidence that Cx43 expression might be restricted to a subpopulation of metastatic tumor cells in close association with the brain vasculature, which supports the notion that brain metastases develop in close proximity to the vasculature.

Metastatic 4T-1 breast cancer cells express Cx43 and form functional GJs with the endothelium. While microtumors develop in association with connexin-rich vasculature and stroma of the brain, the role of connexins in this process has not been examined (Kienast et al., 2010; Lorger and Felding-Habermann, 2010; Carbonell et al., 2009). To investigate this process, we first examined the ability of 4T-1 cancer cells to form GJs and transfer calcein orange dye to a monolayer of cultured EA.hy926 human endothelial cells in vitro (Bauer et al., 1992). 4T-1 is a well-studied mouse breast cancer cell line that readily metastasizes to the
brain and other organs (Serres et al., 2012; Tao et al., 2008; Pulaski and Ostrand-Rosenberg 2001). 4T-1 cells are known to express Cx43 and low levels of Cx26 (Fig. 2A), and they form functional GJs with cultured EA.hy926 cells (Fig. 2B). Importantly, inhibition of Cx43 expression in 4T-1 cells using 3-4 independent Cx43 shRNAs (4T-1KNCx43) (Fig. 2A,B) or siRNA (supplementary material Fig. S2A,B) prevented GJ communication with the endothelium. Interestingly, while loss of Cx43-mediated GJ communication did not impair 4T-1 cell growth under standard adherent culture conditions (Fig. 2C, supplementary material Fig. S2C), it did reduce 3D colony formation and the size of spheroids when cultured alone or co-cultured with endothelial cells (supplementary materials Fig. S3A,B). Similar findings were also obtained using carbenoxolone (CBX), a reported GJ inhibitor (Farina et al., 1998) (Fig. 2B,C, supplementary material Fig. S2A-C, S3A,B). Together these demonstrate that 4T-1 cells form functional Cx43-mediated GJs with endothelial cells in vitro and this process is necessary for spheroid formation and colonization of 3D matrices.

**Cx43 expression is required for metastatic breast cancer cell extravasation and blood vessel co-option in the brain.** We next wanted to know if loss of Cx43 expression would impact brain colonization in preclinical animal models of brain metastasis. In mice, 4T-1KNCx43 cells showed significantly decreased ability to initiate microtumor formation in the brain compared to 4T-1 control cells after 3-7 days post injection into the carotid artery (Fig. 2D). We also used zebrafish and chicken embryo metastasis models since they are more amenable to visualizing initial events that mediate microtumor formation including cancer cell extravasation and vessel co-option using standard multi-color confocal microscopy (Stoletov et al., 2010; Stoletov et al., 2007; Zijlstra et al., 2002). In zebrafish, the majority (>90%) of 4T-1 cells quickly extravasated out of the brain vasculature, and invaded into the surrounding tissue (Fig. 3A,B). In contrast, 4T-1KNCx43 cells displayed significantly less extravasation potential than 4T-1 cells (Fig. 3A,B). Interestingly, 3-D imaging and morphometric shape analyses revealed that thirty-six percent of extravasated 4T-1 cells displayed an elongated invasive shape compared to only eight percent of the 4T-1KNCx43 cells, which displayed a more rounded shape (Fig. 3C). This suggests that connexins may also be involved in mediating shape changes necessary for tissue invasion by breast cancer cells after their extravasation.

The chicken metastasis model is beneficial in that it facilitates visualization and quantification of large numbers of tumor cells that have undergone extravasation and microtumor formation in the brain for up to 7 days post injection. While 4T-1 cells efficiently colonize the chicken brain, both 4T-1KNCx43 cells and 4T-1 siRNA cells depleted of Cx43 show significantly decreased brain colonization (Fig. 3D-F, supplementary material Fig. S2D). Importantly, many of the elongated 4T-1 cells grow along the brain vasculature closely
contacting the vessel wall (Fig. 3D-F). These cells efficiently co-opt pre-existing brain vessels
and form multiple metastatic microtumors, whereas, 4T-1KNCx43 cells show decreased vessel
coop-option, forming less and smaller metastatic lesions (Fig. 3D-F). Together these findings
indicate that Cx43 expression contributes to tumor cell extravasation and microtumor formation
in association with the vasculature.

The metastatic gene twist induces Cx43 expression, tumor cell extravasation, and brain
colonization. Overexpression of the twist transcription factor in breast cancer and melanoma
cells has been reported to increase cell metastasis and correlate with poor patient prognosis
(Yang et al., 2004; Mani et al., 2008; Elenbaas et al., 2001). However, it is not clear how twist
induces tumor cell metastasis in vivo. Importantly, we found that twist overexpression in
HMLE human breast cancer cells (HMLEtwist, Mani et al., 2008) induces increased expression
of Cx43 protein (Fig. 4A,B). This was associated with increased Cx43-dependent GJ coupling
to the endothelium in vitro (supplementary materials Fig. S4A). The depletion of Cx43, or
treatment with CBX did not significantly impact HMLE or HMLEtwist cell proliferation in vitro
(supplementary materials Fig. S4B). These findings demonstrate that expression of the
metastatic gene twist induces Cx43 expression leading to increased GJ communication with
the endothelium.

In zebrafish and chickens, HMLEtwist cells with increased Cx43 expression more
rapidly extravasate and form more microtumors along the vessel surface compared to control
HMLE cells without twist overexpression (Fig. 4A-C). These events required Cx43, as shRNA
mediated knockdown of Cx43 expression in HMLEtwist cells (HMLEtwistKNCx43), inhibited
cell extravasation and brain colonization (Fig. 4C-G). Also, in 4T-1 cells, shRNA mediated
downregulation of endogenous twist expression (a mix of 4-5 independent shRNAs) resulted in
robust relocalization of Cx43 from the cell membrane to the peri-nuclear region (Fig. 5A-C).
This was associated with reduced GJ communication with the cultured endothelium (Fig. 5D).
Inhibition of twist and Cx43 expression or treatment with CBX did not significantly impact 4T-1
cell proliferation in culture (Fig. 5E). Importantly, downregulation of endogenous twist
expression did significantly decrease their ability of to colonize the chicken brain (Fig. 5F).
These data indicate that twist facilitates breast cancer metastasis to the brain by increasing
Cx43 expression and GJ communication with the endothelium.

Metastatic tumor cells form functional heterocellular GJs with the endothelium. The fact
that metastatic breast cancer cells showed increased Cx43-dependent vessel co-option in vivo
and GJ coupling to the endothelium in vitro suggest that successful metastatic cancer cells
form heterocellular gap junctions with the brain endothelium in vivo. To investigate this
possibility, we examined the ability of HMLEtwist and HMLEtwistKNCx43 cells to exchange calcein orange dye with the brain endothelium of live zebrafish (Laird, 2006; Saito-Katsuragi et al., 2007). HMLEtwist cells readily transfer calcein dye to the endothelium, whereas HMLEtwistKNCx43 cells did not indicating that dye transport required functional GJ communication and was not due to passive diffusion or cell lysis (supplementary materials Fig. S5A,B). These findings suggest that cancer cells form functional GJs with brain endothelial cells in a Cx43-dependent manner.

**Metastatic melanoma cells use connexin 26 for brain perivascular microtumor formation.** Similar to breast cancer, melanoma cells often metastasizes to the brain, prompting us to investigate if connexin-mediated tumor cell vessel co-option also plays a role in melanoma brain colonization. For these studies, we used an established mouse metastastic melanoma cell line (b16) that expresses Cx26, but not Cx43, and readily form functional GJs with the cultured endothelium in a Cx26 dependent manner (supplementary materials Fig. S6A,B). b16 cells robustly colonized the chicken brain forming numerous microtumors that invade deep into the tissue along the preexisting vasculature (Fig. 6A). In contrast, b16 cells that were engineered to stably express Cx26-targeting shRNA constructs (b16KNcx26, mix of 4-5 independent shRNAs) formed significantly less metastatic lesions, which were smaller and less invasive (Fig. 6B-D). These results show that in metastatic melanoma cells Cx26 expression enhances microtumor formation in the brain in association with the existing vasculature.

**Survival of dormant, non-proliferating breast cancer cells involves connexin-mediated attachment to the brain vasculature.** A major clinical problem in breast cancer is the inability to eradicate non-proliferating “dormant” breast cancer cells (Nguyen et al., 2009; Chambers et al., 2002). For example, dormant breast cancer cells can reside undetected for many years in the brain before being activated to initiate cancer growth leading to patient relapse (Chambers et al., 2002; Goss et al., 2008). However, since these cells do not proliferate they cannot be targeted by conventional therapies (Goss et al., 2008). While the mechanisms that promote tumor cell dormancy are poorly understood, it is plausible that Cx43-mediated GJ communication with the vasculature is involved in this process (Kienast et al., 2010). Therefore, we investigated the role Cx43 in mediating the survival of dormant breast cancer cells in the brain using the dye retention model of cell dormancy (Kusumbe and Bapat, 2009). To visually monitor dormant breast cancer cells in the brain, GFP expressing 4T-1 breast cancer cells were labeled with PKH26 (red) membrane dye and imaged using confocal microscopy (Fig. 7A). Non-proliferating breast cancer cells retain the dye, while proliferating...
cells rapidly lose the dye after approximately three cell divisions (Kusumbe and Bapat, 2009). Using this approach, we found that the percentage of PKH26 positive 4T-1 cells rapidly declines within the first week post tumor cell injection (from 75% at day 1 post injection to 1.2% at day 7 post injection, Fig. 7B). In contrast, the percentage of PKH26 positive 4T-1 cells that remain in contact with the brain vasculature is increasing within the same period of time showing that non-proliferative dormant breast cancer cells require persistent association with the vasculature to survive (Fig. 7B). In contrast, less PKH positive 4T-1KNcx43 cells were seen contacting vessels in the brain compared to PKH positive 4T-1 cells at day 1 post injection. Importantly, the number of PKH positive 4T-1KNcx43 cells contacting vessels rapidly declined over the 6-day period (Fig. 7B). These data show that non-proliferative dormant metastatic breast cancer cells require contact with the vasculature to survive and that Cx43 is necessary for this process.

**Pharmacological blocking of connexin-mediated cell-cell communication inhibits breast cancer and melanoma brain microtumor formation.** CBX inhibits GJs, pannexins, and hemichannels and is a widely used to treat gastric ulceration and inflammation in humans (Juszczak and Swiergiel, 2009; Farina et al., 1998). It has also been successfully used intracranially in rats and mice to inhibit whole brain GJ communication and to alleviate post-traumatic pain suggesting that CBX could also be used to inhibit GJs in brain metastases (Khorasani et al., 2009; Juszczak et al., 2009; Roh et al., 2010). This and the fact that pharmacological compounds cross the blood-brain barrier in chickens prompted experiments to determine whether CBX treatment inhibited brain colonization and microtumor growth in the chicken brain metastasis model (Ribatti et al., 1993). Strikingly, we found that systemic delivery of CBX, but not vehicle, strongly decreased brain metastatic load of both 4T-1 breast cancer (Fig. 8A-D) and b16 melanoma cells (supplementary materials Fig. S7A-D). CBX treatment also strongly inhibited the ability of 4T-1 breast cancer cells to contact the vasculature resulting in reduced number and size of 4T-1-induced brain microtumors (Fig. 8C,D). Similarly, CBX treatment decreased the number and size of b16 metastatic lesions associated with the vasculature (supplementary materials Fig. S7C,D). Finally, CBX treatment inhibited the size and number of 4T-1 and b16 cells colonies that develop in a reconstituted 3D matrix *in vitro* (supplementary materials Fig. S3A-D). Taken together our findings suggest that connexin-mediated tumor cell-vessel co-option and GJ communication could be a useful therapeutic mechanism to inhibit brain micrometastatic lesion progression.
Discussion

The role of connexins in cancer progression is controversial and not easily reconciled. Most likely this is due to cellular heterogeneity and the complex, multi-step, process of cancer development and progression. A substantial amount of work indicates that Cx43 expression is down regulated in primary breast cancer tumors. It is possible that the loss of GJ expression in primary tumor cells might be important for cells to physically detach from each other. During tumor progression, this could contribute to the local cell invasion into the surrounding stroma and vasculature, which are early steps in the metastatic process (Laird, 2006; Naus and Laird, 2010; Langlois et al., 2010). On the other hand, Cx43 and 26 were found to be up-regulated in established breast cancer and melanoma metastatic lesions suggesting that connexins play roles in late metastatic steps involving vascular penetration and tissue colonization (Elzarrad et al., 2008; Ito et al., 2002; Naoi et al., 2007; Haass et al., 2010; Saito-Katsuragi et al., 2007; Chao et al., 2012). Also, the loss of connexins in primary tumor cells and its reemergence at metastatic sites suggest that connexin expression is dynamically regulated during cancer progression. In this regard, activation of the transcription factor twist is believed to be a late activating event that drives breast cancer and melanoma cell metastasis. We found that twist activation in cancer cells regulates Cx43 expression and its cellular localization, which is necessary for GJ coupling and brain colonization. This and the fact that Cx43 expression was necessary for tumor cell extravasation and microtumor formation is consistent with connexins contributing to late events in the metastatic cascade.

Connexins may also be expressed differentially in primary and disseminating tumor cells. For example, it is possible that connexin expression is restricted to minor populations of tumor cells endowed with invasive and cancer stem cell-like properties (Lawson et al., 2009; Visvader and Lindeman, 2008). These migrating cancer stem cells are believed to be the metastatic culprits that seed secondary tissues. Interestingly, twist has been shown to promote a stem cell-like phenotype in breast cancer cells (Mani et al., 2008). Thus, it is possible that Cx43 expression is due in part to twist activation of a cancer stem cell program. Interestingly, connexin mediated cell-cell adhesion and communication have been shown to be involved in normal stem cell survival and proliferation (Todorova et al., 2008; Cheng et al., 2004; Wong et al., 2006). Also, human and mouse embryonic stem cells as well as human neural progenitors require GJ communication for their survival (Todorova et al., 2008; Cheng et al., 2004; Wong et al., 2006). Consistent with this idea we observed that only minor populations of Cx43-positive metastatic breast cancer cells reside in close proximity to the vascular system in the human brain (Fig. 1). Taken together our findings support the notion that connexins play a later role in regulating cancer progression rather than regulating early events in primary tumor development.
While we show that connexins are required for extravasation and microtumor growth in association with the brain vasculature their precise function in mediating these processes is not completely understood. As part of the extravasation process, connexins may mediate tumor cell adhesion and arrest in the brain vasculature using GJ-independent processes. For example, brain endothelial cell-cell junctions are highly enriched with Cx43, which could serve as homing sites for the attachment and extravasation of tumor cells expressing Cx43. This might explain why breast and melanoma cells commonly metastasize to the brain. Indeed, Cx43 expression is increased in breast cancer cells selected for their propensity to metastasize to the brain (Bos et al., 2009).

It is intriguing that disseminating tumor cells do not completely arrest in the vessel, but migrate along the luminal surface in search of suitable sites to extravasate (Stoletov et al., 2010). Interestingly, Cx43 can regulate cell migration independent of its typical GJ functions (Olk et al., 2009). Increased Cx43-mediated intraluminal cell migration could contribute to the ability of tumor cells to survey and penetrate the vascular wall by forming heterotypic interactions with Cx43-rich endothelial cell-cell junctions.

Our findings suggest that traditional GJ-dependent mechanisms are also likely at play in forming brain metastases. For example, we show that extravasated tumor cells in vivo transfer dye to the brain endothelium in live animals and to cultured endothelial cells in vitro in a Cx43-dependent manner. Also, the GJ communication inhibitor, CBX, potently inhibited dye transfer and perivascular metastatic microtumor formation. These findings suggest that the transfer of ATP, critical ions, small peptides and even small regulatory RNAs through GJ channels could be crucial for tumor cell survival and growth after they leave the circulation (Laird, 2006; Mendoza-Naranjo et al., 2007; Katakowski et al., 2010). In this way, Cx43-mediated extravasation and heterocellular GJ formation with the Cx43-rich brain endothelium could facilitate tumor cell integration into foreign tissues creating a more hospitable niche for metastatic growth. The specific disruption of connexins using pharmaceuticals or genetic approaches could provide an efficient way to eradicate breast cancer and melanoma metastases in the brain.
Materials and Methods

Cell lines, cell culture, siRNA/shRNA constructs, and antibodies

The human mammary carcinoma cell line HMLE and HMLEtwist (GFP or tomato red) were a generous gift from Dr. Jing Yang (UCSD, Yang et al., 2004). Stable fluorescent mouse mammary cancer 4T-1 cells (GFP) were a generous gift from Dr. David Schlaepfer (UCSD) and originally acquired from ATCC. Stable fluorescent b16 (GFP) mouse melanoma cell line was a generous gift from Dr. Brunhilde Felding-Habermann (TSRI) and originally acquired from ATCC. Cells were cultured using standard conditions (www.atcc.org). Cx43 and 26 stable knockdown cell lines (HMLEtwistKNcx43) were generated using commercially available lentiviral vectors (Santa Cruz Biotechnology, catalog # sc-29276-V, sc-35091-V and sc-37051-V). Lentiviral particle production and infection were done according to the manufacturer recommendations. These lentiviral vectors consist of a pool of three to five plasmids designed to achieve maximum knockdown of target gene expression with minimum off target effects when expressed in the same cell. Control cells were established using the empty lentiviral vector. Knockdown was stable through at least 6 passages in culture as determined by western blotting. siRNA knockdown of Cx43 expression in tumor cells was performed using Cx43 siRNAs (sc-29276, according to the manufacture’s instructions. Scrambled siRNA (Santa Cruz Biotechnology) at the same concentration served as a control. Cells were used in experiments 48 hours after transfection. EA.hy926 endothelial cells were acquired from ATCC. Culturex 3D tumor cell-endothelial co-culture was done as previously described (Barkan et al., 2008). Lab-Teck 8 well chamber slides (Fisher) were used for Culturex 3D culture using 2 x 10^4 cells per well for each experiment. For co-culture experiments 1:1 endothelial-tumor cell ratio was used. Cx43 antibody was from Cell Signaling (#3512) and GAPDH antibody was from AbCam (#ab22555).

Oncomine Analyses.

Normalized gene expression signatures were collected using the gene summary viewer for several published breast cancer and melanoma patient arrays to assess the significance of Cx43 and Cx26 gene expression on disease progression and patient outcome. Data was plotted as an average, with a minimum and maximum measurement for each patient group.

Tumor cell to endothelial cell dye transfer.

1 x 10^6/mL tumor cells were preloaded with 30 μM of the cell permeable calcein red-orange dye (#C3481, Life Technologies) for 30 minutes in suspension using standard tissue culture conditions. Cells were then rinsed with PBS and resuspended in culture media (1 x 10^5/mL)
prior to experiments with CBX (10 μM, Sigma) or vehicle. Labeled tumor cells (1,000) were added to a confluent monolayer of EA.hy926 endothelial cells on Lab-Tek 8 well glass bottom chamber slides (Fisher) in the continued presence of 10 μM CBX or vehicle. Cells were allowed to adhere for 30 minutes before dye transfer from tumor cells to the adjacent endothelium was captured using live-cell epifluorescence microscopy. The number of adherent cells that transferred dye to adjacent endothelial cells was recorded and expressed as percent of the total number of tumor cells counted. For each chamber, 40-60 attached cells were recorded. All experiments were repeated twice with triplicate chambers.

Mouse brain metastasis model

Animals were maintained according to the University of California San Diego animal welfare guidelines as described and approved UCSD Institutional Animal Care and Use Committee (S99001). Athymic NCR nude mice (nu/nu), 5-6 weeks of age were used for the study. The breeding pairs were obtained from Charles River Laboratories (Wilmington, MA). Mice were kept in a barrier facility under HEPA filtration and fed with autoclaved laboratory rodent diet. Mice were anesthetized with a ketamine mixture (10 μL ketamine HCL, 7.6 μL xylazine, 2.4 μL acepromazine maleate, and 10 μL H₂O) via s.c. injection. The carotid artery was exposed by blunt dissection and 2.5 x 10⁵ GFP-labeled 4T-1Knx43 or mCherry-labeled 4T-1 parental cells were injected into the right internal carotid artery. The skin was closed with a 6-0 surgical suture (ETHICON, Inc) and 3-7 days later, injected animals were euthanized using CO₂ exposure, their brains were removed and GFP/mCherry positive microtumors localized and counted using using Nikon c1-si confocal microscope, a 10X objective, and Imaris software. At least 3 animals were used for each experiment.

Zebrafish cancer cell extravasation assay

Animals were maintained according to the University of California San Diego animal welfare guidelines as described and approved by UCSD Institutional Animal Care and Use Committee and protocol number S06008 (Nusslein-Volhard and Dahm, 2002). Zebrafish extravasation assay was done as previously described (Stoletov et al., 2010). Briefly, Zebrafish embryos were obtained using standard mating conditions and staged by hours post fertilization (h.p.f). At 48 h.p.f., embryos were de-chorionized and anesthetized with tricaine. 200-300 human tumor cells in 50 nL of PBS were injected into the common cardinal vein using an Eppendorf FemtoJet injector. Embryos that contain brain-arrested cells were imaged using Nikon c1-si confocal microscope, and the percentage of extravasated cells outside the vasculature was quantified as previously described (Stoletov et al., 2010). At least 10 animals were used for each experiment.
Western blotting

Cell extracts were prepared in SDS buffer containing complete protease inhibitor cocktail. Protein concentrations were determined by bicinchoninic acid assay (Pierce). Equal amounts of cell extracts were resolved by SDS-PAGE, electrotransferred to PVDF membranes, and probed with primary antibodies.

Immunofluorescence of cells and human brain breast cancer metastases

Cells were plated on Lab-Tek 8 chamber glass cover slips (Fisher), fixed in 4% formaldehyde, permeabilized in 0.2% Triton X-100, and incubated with anti-Cx43 or ZO-1 antibodies (1:100) followed by secondary antibodies conjugated with Alexa Fluor 488 or Alexa Fluor 594 (Molecular Probes). Coverslips were mounted using ProLong Gold (Invitrogen) mounting media and imaged using a Nikon C1-si confocal microscope. Images were obtained using a 60X oil immersion objective and analyzed using Imaris imaging software.

Breast cancer brain metastasis tissue samples from four patients were obtained from UCSD Moores Cancer Center Tissue Repository. Tissue sections were cut from blocks of formalin-fixed paraffin tumor tissue of UCSD patients that had metastatic breast cancer to the brain. Four-micron tissue sections were stained with a rabbit polyclonal antibody to Cx43 (#3512, Cell Signaling Technology, Inc), and a mouse monoclonal antibody to GFAP (E16510, Spring Bioscience) using Discovery Ultra (Ventana Medical Systems). Antigen retrieval was performed using CC1 for 24 minutes at 95 °C. Primary antibodies were used at 1:150 dilution for Cx43 and 1:600 dilution for GFAP for 1 hour at 37 °C followed by fluorescently labeled (donkey anti-rabbit 594; donkey anti-mouse 488) secondary antibodies (Jackson ImmunoResearch) at 1:750 dilution for 60 minutes. Slides were rinsed, and coverslipped (Prolong Gold with DAPI, Invitrogen). Slides were visualized on a Zeiss Axio Imager2 using Cambridge Research Instruments (CRi) Nuance Multispectral Imaging System software to capture the images and unmix pure spectra. Using this software, a pure fluorophore spectrum is acquired and then compared to a complex mixture of spectrum, which includes autofluorescence. Individual fluorophore spectrum as well as autofluorescence can be individually displayed.

Chicken embryo cancer metastasis assay

Fertilized chicken eggs were purchased from McIntire Poultry (Lakeside, CA) and incubated for 14 days (37°C, 100% humidity). A 1cm² square window was created in the eggshell to expose the underlying vasculature. Tumor cells were suspended in sterile PBS at the concentration of 2 x 10⁶/ml and 2 x 10⁵ tumor cells (100 µl/embryo) were injected into the main CAM vein. After incubating embryos for another 1-7 days, Rhodamine-lectin (Vector Biolabs, 100 µL/embryo,
0.5 mg/mL) was injected into the CAM vein. Following appropriate euthanizing procedures, the
chicken brain and liver were removed and imaged using either Leica MZFLIII microscope
(1.25X) or Nikon c1-si confocal microscope (10-60X). CBX (10 mg/mL in PBS, (Sigma) was
injected into the chicken embryo amniotic fluid 2.5 mg/embryo/day. CBX treatment led to
approximately 30% decrease in the embryo survival however no gross organ or tissue
morphology effects were observed. Cancer dormancy and PKH membrane dye (Sigma)
staining was performed as according to the manufactures recommendation and as previously
described (Kusumbe and Bapat, 2009).

Statistical analysis
All data were analyzed using unpaired Student’s t-test built into the GraphPad Prizm software
(www.graphpad.com) for statistical significance. Data plots show mean values ± s.e.m.

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References


Elenbaas, B., Spirio, L., Koerner, F., Fleming, M.D., Zimonjic, D.B., Donaher, J.L.,


**Figure Legends**

**Fig. 1. Cx43 expression pattern in brain metastases from breast cancer patients.**
Epifluorescent photomicrographs of a surgically resected brain metastasis from a breast cancer patient triple stained for (A) nuclei (DAPI, blue), (B) Cx43 (red), and (C) the glial cell marker protein GFAP (green). (D) shows the merged images of A-C. (E) zoomed region of box in (D). Asterisk in (A) and box in (D) show vessel lumen with autofluorescent red blood cells (red, arrowhead). Asterisk in (E) shows tumor on the vessel surface with positive Cx43 staining. (F-J) Epifluorescent photomicrographs of uninvolved normal brain tissue and associated blood vessel from the same brain metastasis specimen as above. (J) zoomed region of box in (I). Asterisk in (F) and box in (I) show the vessel lumen with autofluorescent red blood cells (red, arrowhead). Bars = 25 μm in A, F, and 10 μm in E,J.

**Fig. 2. Inhibition of Cx43 expression in breast cancer cells inhibits GJ communication *in vitro* and inhibits brain colonization in mice.**
(A) 4T-1 cells were either treated with an empty lentiviral vector (Control) or treated with the lentiviral vector encoding shRNA to Cx43 (4T-1KNcx43) to knockdown Cx43 expression. Stable cells lines were then selected and Cx43 expression levels examined by western blotting. Actin, GAPDH, and Cx26 served as specificity and loading controls. 4T-1KNcx43 cells show a 78% decrease in Cx43 expression compared to 4T-1 control cells, as measured by densitometry. (B) The indicated tumor cells were prelabeled with calcein orange dye and then added to a monolayer of EA.hy926 endothelial cells in the presence of the GJ inhibitor CBX (10 μM) or its vehicle PBS. Dye transfer from tumor cells to endothelial cells was observed live by epifluorescence microscopy after 30 minutes of co-culture. The number of adherent cells that transferred dye to the adjacent endothelium was determined and represented as percent of total number of tumor cells counted. * = P values < 0.05 (C) The indicated tumor cells were cultured *in vitro* and examined for cell growth for 3 days in the presence of CBX (10 μM) or its vehicle using the CyQUANT assay. rfu = relative florescence units. P >0.05 (D) Average number of micrometastatic lesions in the mouse brain induced by 4T-1 and 4T-1KNcx43 cells, 3-7 days post injection * = P values < 0.05.

**Fig. 3. Metastatic breast cancer cell extravasation and brain colonization require Cx43 expression.**
(A) 4T-1 control or 4T-1KNcx43 cells expressing dsRed were injected into the circulation of Tg(fli1:egfp) transgenic zebrafish and allowed to extravasate in the brain for 24 hours. Shown are representative 3D confocal reconstructions of the brain vasculature (green) and tumor cells
(red) in the head region. (B) Average percentage of 4T-1 control and 4T-1KNCx43 cells that extravasated out of the vasculature into the brain parenchyma 24 hours after injection. (C) Average percentage of extravasated 4T-1 control and 4T-1KNCx43 cells with an elongated invasive morphology in the brain parenchyma. Bars, 200 µm. (D) 4T-1 control or 4T-1KNCx43 cells expressing GFP were injected into the chicken circulation and allowed to form microtumors in the brain for 7 days. Shown are representative 3D confocal reconstructions of the brain vasculature (red, Rodamine-lectin stained) and tumor cells (green)(60X). Insets show lower magnification (10x) of areas enlarged in the main panels delineated by the dashed squares. Note that 3D rendering shows that 4T-1 control cells co-opt brain blood vessels while 4T-1KNCx43 show little vessel association. (E) Number and volume of 4T-1 control and 4T-1KNCx43 microtumor lesions in the chicken brain after 7 days. (F) Average length of tumor-cell blood vessel contacts for 4T-1 control and 4T-1KNCx43 tumor cells after 7 days in the chicken brain. Bars, 20 µm (main panels) or 200 µm (insets).

**Fig. 4. Expression of the metastatic gene *twist* in breast cancer cells induces Cx43 expression, tumor cell extravasation, and microtumor formation in the brain.**

(A) Representative images show 3D reconstructions of confocal z-stacks (60x, top views) of HMLE or HMLEtwist cell monolayers co-immunostained with connexin 43 (Cx43, green) and the cell-cell junction marker zonula occluden-1 (ZO-1, red). (B) Western blots of the indicated proteins in HMLE cells transduced with the empty lentiviral vector (HLME), transduced with lenti-twist (HMLEtwist) or expressing twist and shRNAs to Cx43 (HMLEtwistKNCx43). HMLEtwist cells showed a 2.3 fold increase in Cx43 compared to HLME cells and HMLEtwistKNCx43 cells showed a 2.0 fold decrease compared to HMLEtwist cells, as measured by densitometry. (C) HMLE, HMLEtwist or HMLEtwistKNCx43 cells expressing dsRed were injected into the circulation of Tg(*fli1:egfp*) transgenic zebrafish and allowed to extravasate in the brain for 24 hours. Shown are representative 3D confocal reconstructions (20X) of the brain vasculature (green) and tumor cells (red) in the head region. The left most panel shows 3D reconstruction of the head of a control, non-injected embryo. Insets show higher magnification (60x) of areas delineated by the dashed squares. (D) Brain microtumor formation of HMLE, HMLEtwist and HMLEtwistKNCx43 cells was assessed using the chicken metastasis assay. Representative 3D reconstructions (10X, top views) show confocal z-stacks of chicken embryo brains 3 days after intravenous injection of HMLE, HMLEtwist or HMLEtwistKNCx43 cells. (E) Average percent of HMLE, HMLEtwist or HMLEtwistKNCx43 cells that extravasated in the zebrafish embryo brain 24 hours after injection. (F) Average number of microtumors per zebrafish brain induced by HMLE, HMLEtwist and HMLEtwistKNCx43 cells 24 hours after injection. (G) Average number of microtumors per chicken brain induced by HMLE,
HMLEtwist and HMLEtwistKNCx43 cells 3 days after injection. Bars, 20 µm (A) or 200 µm (C, D).

**Fig. 5. Twist inhibition in metastatic breast cancer cells reduces Cx43-mediated GJ communication and inhibits brain microtumor formation.** (A) Representative images show 3D reconstructions of confocal z-stacks (60x, top views) of 4T-1 expressing the empty lentiviral vector or 4T-1 cells expressing lentiviral vectors encoding shRNAs to *twist* (4T-1KN-twist). Cells were co-immunostained for Cx43 and ZO-1 and imaged as in figure 4A. Note that in cells with *twist* depleted Cx43 is mostly intracellular, whereas, ZO-1 is localized to the membrane. (B) Western blots of indicated proteins in 4T-1 and 4T-1KN-twist cells. Note that loss of twist expression inhibits membrane localization of Cx43 as shown in (A), but not Cx43 total protein levels. 4T-1KN-twist cells show a 71% decrease in Twist1 expression compared to 4T-1 cells, as measured by densitometry. (C) Pearson’s correlation analyses of Cx43 and ZO-1 co-localization in 4T-1 and 4T-1KN-twist cells immunostained as in (A). * = *P* values < 0.05. Bar, 20 µm. (D) 4T-1 and 4T-1KN-twist cells in the presence of CBX (10 µM) or vehicle were examined for GJ-mediated calcein orange dye transfer to the cultured endothelium as described in figure 2B. * = *P* values < 0.05 (E) 4T-1 and 4T-1KN-twist cell growth in the presence of CBX (10 µM) or vehicle was determined as in figure 2C. *P* >0.05. (F) The number of 4T-1 and 4T-1KN-twist microtumors was assessed after 7 days in the chicken brain as described in figure 3D. * = *P* values < 0.05.

**Fig. 6. Metastatic melanoma brain colonization requires Cx26** (A) b16 metastatic melanoma cells expressing the empty lentiviral vector (b16 control) or (B) b16 cells stably expressing lentiviral shRNAs to Cx26 (b16KNCx26) along with GFP were injected into the chicken circulation and allowed to form microtumors in the brain for the indicated days. Left two panels are representative 3D confocal reconstructions of the brain vasculature (red, Rodamine-lectin stained) and tumor cells (green)(60X). Insets show lower magnification (10x) of areas enlarged in the main panels delineated by the dashed squares. Note that 3D rendering shows that b16 control cells co-opt brain blood vessels while b16KNCx26 cells show little vessel association. Middle panels show confocal z-stacks as side views. Right panels show representative optical slices from the middle of the z-stacks (slice positions indicated by the dashed lines in the middle panels). (C) Representative whole chicken brain images 4 days after intravenous injection of either b16 or b16KNCx26 cells showing the level of brain colonization of GFP labeled tumor cells. (D) Average volume and number of brain microtumors induced by b16 mock or b16KNCx26 cells 4 days after injection. Bars, 200 µm (A, B) or 0.5 cm (C).
Fig. 7. Cx43 expression is necessary for survival of non-proliferating dormant breast cancer cells in the brain.
(A) 4T-1 cells expressing GFP (green) and stained with the dye PKH26 (red) were injected into the chicken circulation and allowed to form microtumors in the brain for the indicated days. Confocal images were captured as described in figure 3D. Non-proliferating, dormant tumor cells, retain PKH26 and appear yellow with the green GFP background. The chicken brain vasculature is labeled with Rhodamine-lectin (red). Insets show higher magnification of areas delineated by the dashed squares. Representative example of PKH26 negative microtumor at day 7 post tumor cell injection (day 7, lower right corner). (B) Average percentages of surviving PKH26 positive 4T-1 cells (green line) or PKH26 positive 4T-1 and 4T-1KNcx43 cells that directly contact blood vessels in the chicken brain for the indicated days (red line, and black line, respectively). Bar, 200 µm.

Fig. 8. The GJ inhibitor CBX inhibits breast cancer brain colonization.
(A) Chickens were treated with vehicle or (B) CBX (2.5 mg/egg/day) starting one day before tumor cell injection. Microtumor formation in the brain was assessed at the indicated days using confocal microscopy as described in figure 6B. Insets show higher magnification (60x) of the areas outlined by the dashed squares. The brain vasculature is labeled with Rhodamine-lectin dye (red). (C) Average length of tumor-cell blood vessel contacts for 4T-1 cells from the brains of vehicle or CBX-treated chicken 6 days after injection. (D) Number and volume of microtumors per chicken brain induced by 4T-1 tumor cells for vehicle or CBX treated chickens 6 days after injection. Bars = 200 µm.

Supplemental Figure Legends:

Fig. S1. Oncomine analyses show positive correlation of Cx43 and Cx26 expression in primary tumors with poor breast cancer and melanoma patient prognosis. (A) Correlation of patient survival outcome with Cx43 expression in primary breast cancers (5 years post diagnosis, Desment dataset). (B) Clinical correlation of cancer recurrence after 3 years with Cx43 expression in primary breast cancers (Finak dataset). (C-E) Correlation of increased primary tumor Cx43 expression with increased risk of melanoma metastasis (1-5 years post diagnosis, Laurent melanoma dataset). (F) Correlation of increased Cx43 primary melanomas with decreased patient survival (5 years post diagnosis, Bittner melanoma dataset). (G) Correlation of increased Cx26 expression in the primary melanomas with decreased patient survival.
survival (5 years post diagnosis, Bittner melanoma dataset). (H) Correlation of increased Cx26 expression in the primary breast cancer stroma tissue with breast cancer recurrence (5 years post diagnosis, Finak breast cancer dataset).

Fig. S2. siRNA depletion of Cx43 inhibits GJ communication, and microtumor formation in the brain. (A) 4T-1 cells were treated with a scrambled control RNA (Scr) or siRNA to Cx43 (siRNAcx43) and then western blotted for the indicated proteins. (B) The indicated tumor cells were prelabeled with calcein orange dye and then added to a monolayer of EA.hy926 endothelial cells in the presence of the GJ inhibitor CBX (10 μM) or its vehicle (PBS). Dye transfer from tumor cells to endothelial cells was observed live by epifluorescence microscopy after 30 minutes of co-culture. The number of adherent cells that transferred dye to the adjacent endothelium was determined and represented as percent of total number of tumor cells counted. * = P values < 0.05 (C) The indicated tumor cells were cultured in vitro and examine for cell growth for 3 days in the presence of CBX or its vehicle using the CyQUANT assay. rfu = relative florescence units. (D) 4T-1scr or 4T-1siRNAcx43 cells expressing GFP were injected into the chicken circulation and allowed to form microtumors in the brain. The number of microtumor lesions per 10X field in the brain was determined by confocal microscopy 5 days after injection. * = P values < 0.05

Fig. S3. Blocking GJ communication inhibits endothelial cell-induced breast cancer and melanoma colony formation in vitro.
(A) 4T-1 tumor cells expressing GFP (green) were cultured for 5 days in three-dimensional Culturex matrix alone or with human EA.hy926 endothelial cells (red, RFP transfected) and treated with vehicle (DMSO) or CBX (20 μM). Upper panel shows 4T-1 cells that were cultured alone and vehicle treated (days 1 and 5) or treated with CBX (day 5). Lower panel shows 4T-1 cells that were co-cultured with EAhy.926 and vehicle treated (days 1 and 5) or treated with CBX (day 5). Note, that co-culture with EA.hy926 cells promotes 4T-1 tumor cell colony formation, while CBX treatment inhibits it. (B) Average size and number of 4T-1 or 4T-1KNcx43 tumor cell colonies that were cultured alone or with EA.hy926 cells and treated with CBX or vehicle (control) for 5 days. (C) b16 melanoma cells expressing GFP (green) were cultured for 5 days in 3D Culturex matrix alone or with human EA.hy926 (red, RFP) endothelial cells and treated either with the vehicle (DMSO) or CBX (20 μM). Upper panel shows b16 cells that were cultured alone and vehicle treated (days 1 and 5) or treated with CBX (day 5). Lower panel shows b16 cells that were co-cultured with EA.hy926 and vehicle treated (days 1 and 5) or treated with CBX (day 5). Note, that co-culture with EA.hy926 cells promotes b16 tumor cell colony formation, while CBX treatment inhibits it. (D) Average size and number of b16 or
b16KNcx26 cell colonies cultured alone or with EA.hy926 and treated with CBX or vehicle (control) as above. Bars = 200 µm. * = P values < 0.05.

**Fig. S4. Twist expression induces breast cancer Cx43-mediated GJ communication with the endothelium in vitro.**

(A) HMLE cells transduced with a empty lentiviral vector (HMLE), transduced with lenti-twist (HMLEtwist) or expressing twist along with lentiviral shRNAs to Cx43 (HMLEtwistKNCx43) were tested for their ability to transfer calcein orange dye to the endothelium as described in Fig. S2B. (B) The indicated HMLE cells were cultured *in vitro* and examined for cell growth for 3 days in the presence of CBX (10 µM) or its vehicle using the CyQUANT assay. rfu = relative florescence units. * = P values < 0.05. ** = P values < 0.01.

**Fig. S5. HMLEtwist cells exchange calcein orange dye with brain endothelial cells in connexin 43-dependent manner in vivo.**

(A) HMLEtwist or (B) HMLEtwistKNcx43 cells were preloaded with calcein orange dye (shown as blue) and injected into the circulation of zebrafish and allowed to form microtumors along the vasculature (green). Dye transfer to the endothelium was monitored in live animals using confocal microscopy. Left panels show 3D reconstructions (20x) of the zebrafish head region with seeded tumor cells. Right panels show higher magnification (60x) of tumor cells delineated with dashed squares. Note the strong presence of the calcein orange dye within the endothelial cells (white arrows) that are located next to HMLEtwist cells, but not in the endothelium adjacent to HMLEtwistKNCx43 cells. Bars = 100 µm in A and 20 µm in B.

**Fig. S6. Inhibition of Cx26 expression in b16 melanoma cells inhibits GJ communication with the endothelium.**

(A) b16 metastatic melanoma cells expressing the empty lentiviral vector (b16 control) or b16 cells stably expressing lentiviral shRNAs to Cx26 (b16KNcx26) were western blotted for the indicated proteins. (B) b16 control and b16KNcx26 cells were tested for their ability to transfer calcein orange dye to the endothelium as described in Fig. S2B. * = P values < 0.05 (C) The indicated tumor cells were cultured *in vitro* and examine for cell growth after 3 days in the presence of CBX or its vehicle using the CyQUANT assay. rfu = relative florescence units.

**Fig. S7. The GJ inhibitor CBX inhibits melanoma brain colonization.**

(A) Chickens were treated with vehicle or (B) CBX (2.5 mg/egg/day) starting one day before the tumor cell injection. b16 microtumor formation in the brain was assessed at the indicated days using confocal microscopy as described in figure 6B. Insets show higher magnification
(60X) of the areas outlined by the dashed squares. The brain vasculature is labeled with Rhodamin-lectin (red). Two left panels show the 3D reconstructions of confocal z-stacks (top views), middle panels show the side views of confocal z-stacks. Right panels show representative optical slices from the middle of the z-stacks (slice positions indicated by the dashed lines in the middle panels). (C) Representative whole brain images 4 days after intravenous injection of b16 cells expressing GFP (green). (D) Number and average volume of b16 microtumors per brain in vehicle or CBX treated embryos 4 days after injection. Bars = 20 \mu m in A, B and 0.5 cm in C.
Figure 1A-J
Figure 3A-F

A

4T-1 Control

4T-1 KNcx43

Brain

Eye

B

% Extravasated cells

4T-1 Control

4T-1 KNcx43

p<0.05

C

% Elongated cells

4T-1 Control

4T-1 KNcx43

p<0.05

D

4T-1 Control

Day 7

4T-1 KNcx43

Day 7

E

Microtumors/brain

4T-1 Control

4T-1 KNcx43

p<0.05

F

Microtumor volume µm³ x 10³

4T-1 Control

4T-1 KNcx43

p<0.05

Tumor cell/vessel contact µm

4T-1 Control

4T-1 KNcx43

p<0.05
Figure 5A-F

A

IF: Cx43 ZO-1

4T-1

4T-1KNtwist

B

Twist1
Cx43
GAPDH

4T-1
4T-1KNtwist

C

Pearson's correlation

ZO-1/Cx43

4T-1
4T-1KNtwist

p<0.05

D

% Dye Coupling

4T-1
4T-1KNtwist

E

Cell Number (rfu)

4T-1
4T-1KNtwist

F

Microtumors (Brain)

4T-1
4T-1KNtwist
Figure 7A & B

A chick brain vasculature 4T-1 cells PKH 4T-1 positive cells

B

% Surviving PKH+ cells

Days 0 1 2 3 4 5 6 7

% PKH cells
% PKH cells with vessel contact
% KNcx43 PKH cells with vessel contact
Figure 8A-D

Chicken brain vasculature 4T-1 cells

Top view  Side view  Middle view

Day 1  Day 7

Vehicle

CBX, 2.5mg/egg/day

Cell/Wessel Contact (µm)

Vehicle  CBX

p<0.05

D

Microtumors/brain

*  p<0.05

Microtumors Volume (µm³ x 10⁵)

*  p<0.05

Vehicle  CBX